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7 **Paper Title**

8 Urinary angiotensinogen level is increased in preterm neonates

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19 **Full Names of Authors**

20 Masashi Suzue, Maki Urushihara, Ryuji Nakagawa, Takahiko Saijo, Shoji Kagami

21 Department of Pediatrics, Institute of Health Biosciences, The University of Tokushima

22 Graduate School, Tokushima, Japan

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46 **Concise Title**

47 Urinary angiotensinogen during renal development in neonates

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65 **Corresponding Author**

Maki Urushihara, MD, PhD

Assistant Professor, Department of Pediatrics

Institute of Health Biosciences, The University of Tokushima Graduate School

Kuramoto-cho 3-18-15, Tokushima, Tokushima 770-8503, Japan

Tel: +81-88-633-7135, Fax: +81-88-631-8697

E-mail: urushihara@tokushima-u.ac.jp

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Abstract

Background

All components of the renin-angiotensin system (RAS) are abundantly synthesized in the developing kidney, suggesting that the RAS plays an important role in renal development.

To examine this system in human neonates, we measured urinary angiotensinogen levels in preterm and full-term neonates, and examined the relationship between urinary angiotensinogen levels and gestational age.

Methods

Urine and plasma samples were collected from 20 preterm and 18 full-term neonates at birth.

Angiotensinogen levels were measured using enzyme-linked immunosorbent assay.

Results

Plasma angiotensinogen concentrations were not increased in preterm neonates compared to that in full-term neonates ($P = 0.7288$). However, the urinary angiotensinogen-to-creatinine ratio was significantly higher in preterm neonates compared to that in full-term neonates ($P = 0.0011$). Importantly, the urinary angiotensinogen-to-creatinine ratio dropped significantly with increasing gestational age ($P = 0.0010$), whereas the plasma angiotensinogen concentration was not correlated with gestational age ($P = 0.7814$).

Conclusions

These results suggest that urinary angiotensinogen levels may indicate the involvement of intrarenal RAS activation in prenatal renal development.

Keywords

Renin-angiotensin system; neonates; renal development; gestational age; preterm

Introduction

1
2 The role of the renin-angiotensin system (RAS) in blood pressure regulation and sodium and
3 fluid homeostasis is well recognized. The focus of interest regarding the RAS has shifted to
4 the investigation of its role in specific tissues [1, 2]. The mechanism of the tissue RAS in
5 the kidney is unique because the components necessary to generate Ang II are produced
6 within the kidney [3]. Locally produced Ang II induces inflammation, cell growth
7 mitogenesis, apoptosis, migration, and differentiation, regulates the gene expression of
8 bioactive substances, and activates multiple intracellular signaling pathways [4].

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19 A large number of studies have shown that the RAS plays a key role in the development
20 of the mammalian kidney [5]. Recent studies have addressed the importance of an intact
21 RAS cascade during postnatal kidney development using both pharmacological inhibition and
22 genetic deletion of various RAS components. The mechanisms by which RAS supports
23 renal development are not fully understood, but the temporal and spatial expression of
24 different components of the system suggests a direct action on receptors expressed in the
25 developing structures of the immature kidneys [5].

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36 Angiotensinogen (AGT) is the only known substrate for renin that is a rate-limiting
37 enzyme of the RAS. Recent studies showed that urinary excretion rates of AGT provided a
38 specific index of intrarenal RAS status [6, 7]. A direct quantitative method for measuring
39 urinary AGT using human AGT enzyme-linked immunosorbent assays (ELISA) has been
40 developed [7]. This method demonstrated significantly increased urinary AGT levels in
41 pediatric chronic glomerulonephritis patients who were not treated with RAS blockers
42 compared to those in control subjects [8]. Moreover, the urinary AGT level was
43 significantly higher in pediatric patients with type 1 diabetes in the pre-microalbuminuric
44 phase compared to that in control subjects [9]. However, urinary AGT level in neonatal
45 period is not yet completely documented. These data prompted us to measure urinary AGT

1 in neonates and investigate correlations with clinical parameters. Therefore, this study was
2 performed to test the hypothesis that urinary AGT is a biomarker of intrarenal RAS status
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4 during renal development.
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9 **Methods**

10 *Patients and Urine Samples*

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12 The study's experimental protocol was approved by the Institutional Review Board of
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14 Tokushima University (approval number; 1425). Study participants were recruited in
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16 Tokushima University between April 1, 2012 and March 31, 2013. Neonates were excluded
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18 if they had known or suspected sepsis, severe respiratory distress syndrome, congenital heart
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20 disease, or a renal or chromosomal abnormality. Neonates expected to die within 48 hours
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22 of recruitment were also excluded. Informed consent was obtained from the parents.
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24 Demographic-perinatal characteristics including gestational age (GA), birth weight, mode of
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26 delivery, sex, and Apgar scores at 1 and 5 min were recorded for all neonates. A random
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28 spot urine and a blood samples were obtained in the few days following birth. Urine
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30 samples for analysis were obtained by collecting the urine into urine bags. The urine was
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32 stored at -20°C until biochemical analysis.
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43 *Measurements*

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45 Urinary concentrations of creatinine were measured using the Creatinine Assay Kit
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47 (BioVision, Milpitas, CA). Plasma and urinary concentrations of AGT were measured using
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49 commercially available ELISA kits (IBL, Takasaki, Gunma, Japan), as previously described
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51 [8]. The sensitivity of this assay is >0.31 ng/mL. The intra- and inter-assay coefficients of
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53 variation were 4.4% and 4.3%, respectively. The urinary level of AGT was expressed in
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55 mg/g creatinine (Cr). Plasma Cystatin C measurements were performed by ELISA
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2 according to the manufacturer instruction (R&D, Minneapolis, MN), with intra- and
3 inter-assay coefficients of variation <5.9%.

4 5 6 7 *Statistical Analysis*

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9 Pearson correlation coefficients and Spearman correlation coefficients were used for
10 parametric data and nonparametric data, respectively. Unpaired *t* test was performed to
11 determine the group means. All data are presented as means \pm standard error of mean
12 (SEM). A P-value <0.05 was considered statistically significant. All computations,
13 including data management and statistical analyses, were performed using JMP software
14 (SAS Institute, Candler, NC) and GraphPad Prism software (GraphPad Software, Inc., La
15 Jolla, CA, USA).
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28 **Results**

29 *Subject Profiles*

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31 The profiles of the study participants are summarized in Table 1. A total of 38 neonates,
32 including 20 preterm and 18 full-term neonates, were recruited. The average GAs of
33 preterm and full-term neonates were 32.0 ± 0.6 weeks and 37.7 ± 0.2 weeks, respectively.
34 Preterm and full-term neonates weighed $1,757 \pm 120$ g and $2,611 \pm 131$ g, respectively.
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36 There were no significant differences between preterm and full-term neonates in delivery
37 process, gender, Apgar score at 5 min, the days of sampling after birth, and plasma Cystatin C
38 level. The average Apgar score at 1 min was significantly lower in preterm than in full-term
39 neonates.
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56 *Urinary and Plasma AGT Levels in Preterm Neonates*

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58 The urinary AGT-to-creatinine ratio (Figure 1A) was significantly increased in preterm
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1 neonates (3.58 ± 0.57 mg/g Cr) compared to that in full-term neonates (1.26 ± 0.26 , P =
2 0.0011). However, an increase in plasma AGT levels was not observed (Figure 1B).
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6 7 *Single Regression Analysis*

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9 The urinary AGT-to-creatinine ratio (Figure 2A) was significantly inversely correlated with
10 gestational age ($r = 0.5115$, $P = 0.0010$). However, the plasma AGT level (Figure 2B) was
11 not correlated with gestational age ($r = 0.0465$, $P = 0.7814$).
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19 **Discussion**

20 We compared the urinary AGT-to-creatinine ratio between preterm and full-term neonates.

21 The urinary AGT-to-creatinine ratio was significantly higher in preterm neonates compared to
22 that in full-term neonates. Notably, an increase in plasma AGT levels was not observed.
23

24 Furthermore, the urinary AGT-to-creatinine ratio was inversely correlated with the gestational
25 age. These data suggest that intrarenal RAS activation leads to renal development in the
26 earlier gestation period and that increased urinary AGT levels in preterm neonates reflect this
27 activation.
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29 In human embryos, all components of the RAS are expressed in the kidney at as early as
30 5 weeks of gestation [10, 11]. Their production is precisely time-regulated, suggesting that
31 Ang II could also exert its effects as a growth-promoting agent during kidney development
32 [12] and levels of circulating renin and Ang II are higher during fetal life than during
33 postnatal life [11]. During the gestation, the RAS of the fetal lamb responds to the same
34 stimuli, such as blood volume depletion, furosemide, hypoxemia, and RAS blockade [13, 14].
35 In the same way, human fetuses exposed in utero to RAS blockers are severely hypotensive at
36 birth, and sometimes develop irreversible renal lesions responsible for renal failure and anuria
37 [15, 16]. On the other hand, inappropriate activation of the RAS during fetal life may have
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deleterious consequences [17]. Thus, RAS plays an important role in kidney development.

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2 Recently, multiple studies resulted in the accumulation of evidence indicating that
3 urinary AGT can be a useful tool for identifying the activated intrarenal RAS in hypertension,
4 diabetic nephropathy, and chronic kidney disease [2, 18]. We previously reported that
5 urinary AGT levels are increased in pediatric patients with chronic glomerulonephritis and
6 that treatment with RAS blockers suppressed urinary AGT levels [8], and increased urinary
7 AGT levels precede increased urinary albumin levels and may be one of the earliest predictors
8 of intrarenal RAS activation in juvenile type 1 diabetes [9]. These findings suggest that
9 urinary AGT can be a novel biomarker of intrarenal RAS activation in pediatric patients.
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11 These data along with the present study suggest that urinary AGT levels may reflect intrarenal
12 RAS activation associated with kidney development in neonates.
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26 Although most circulating AGT is produced and secreted by the liver, the AGT
27 produced in proximal tubule cells appears to be secreted directly into the tubular lumen, in
28 addition to producing its metabolites intracellularly and secreting them into the tubular lumen
29 [19]. Proximal tubular AGT concentrations in anesthetized rats have been reported to be in
30 the range of 300–600 nM, which greatly exceeds free Ang I and Ang II tubular fluid
31 concentrations [3]. Because of its molecular size (50–60 kDa), it seems unlikely that much
32 of the plasma AGT filters across the glomerular membrane, further supporting the concept
33 that proximal tubular cells secrete AGT directly into the tubules [20]. To determine if
34 circulating AGT is a source of urinary AGT, the glomerular permeability of AGT was
35 examined by multiphoton imaging [21]. The glomerular permeability of injected exogenous
36 AGT was extremely low. By contrast, urinary excretion of endogenous AGT was
37 significantly increased. These results indicate that the majority of urinary AGT originates
38 from the kidney rather than from glomerular filtration [21]. Consistent with this concept,
39 plasma AGT levels did not differ between preterm and full-term neonates despite significant
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1 between-group differences in urinary AGT levels in the present study. Therefore, it seems
2 highly likely that AGT in urine originates from AGT in the kidney, not from AGT in plasma.
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4 Taken together, our findings reveal physiological significance that measuring urinary AGT
5 in neonate is noninvasive and useful tool for the evaluation of RAS activation during kidney
6 development independent of systemic RAS.
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11 The relatively small sample size in this study is a potential limitation. Furthermore,
12 the study was cross-sectional, and therefore it might be difficult to draw any causal
13 conclusion. However, our observation demonstrates that levels of urinary AGT are increased
14 in preterm neonates compared to that in full-term neonates, suggesting intrarenal RAS
15 activation during kidney development. This pilot study triggered new insight into the
16 mechanism of kidney development that requires further prospective investigation in larger
17 multicenter studies.
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30 study was supported by JSPS KAKENHI Grant Numbers 23591569 and 23591570.
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34 **Conflict of interest**

35 The authors declare no conflict of interests.
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Figure Legends

Figure 1.

Urinary AGT-to-creatinine ratio (Urinary AGT/Cre) and plasma AGT level in preterm and full-term neonates. The urinary AGT/Cre was significantly greater in preterm neonates compared to that in full-term neonates (A). Importantly, there was no difference in plasma AGT levels between these groups (B). Line at mean with SEM.

Figure 2.

Single regression analyses for urinary AGT-to-creatinine ratio (Urinary AGT/Cre) with gestational age (A) and plasma AGT levels with gestational age (B). The urinary AGT/Cre showed inverse correlations with gestational age, but plasma AGT levels did not. Two curves (dashed) surrounding linear regression line (solid) define the 95% confidence interval.

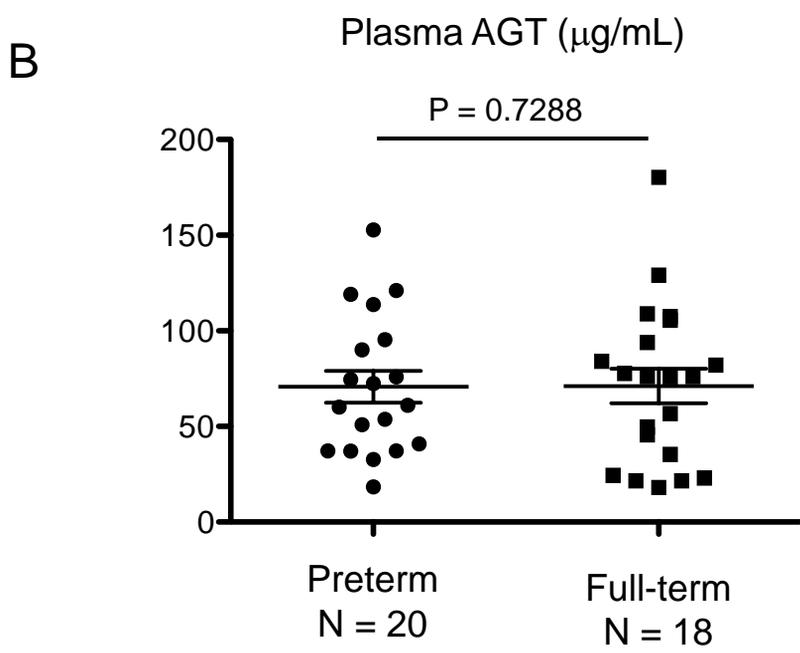
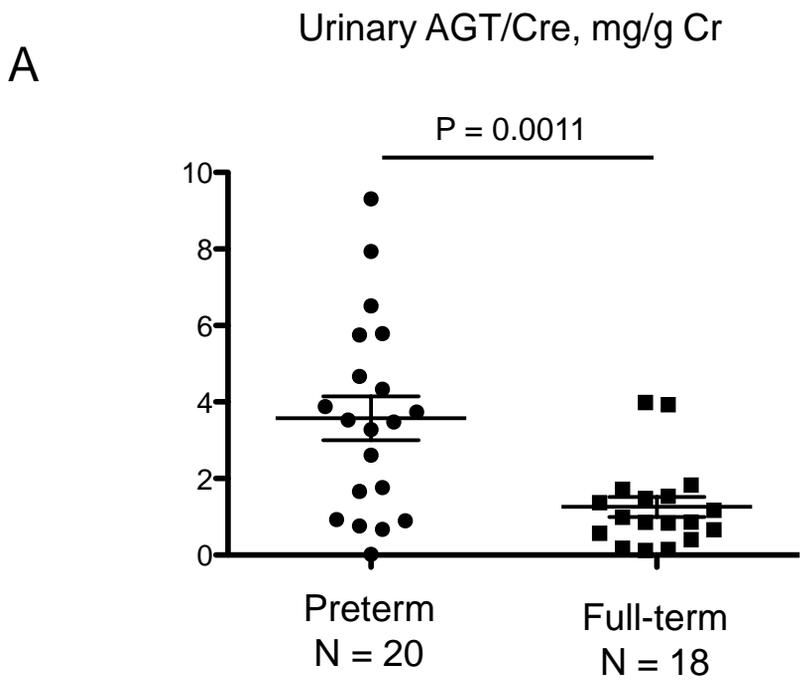
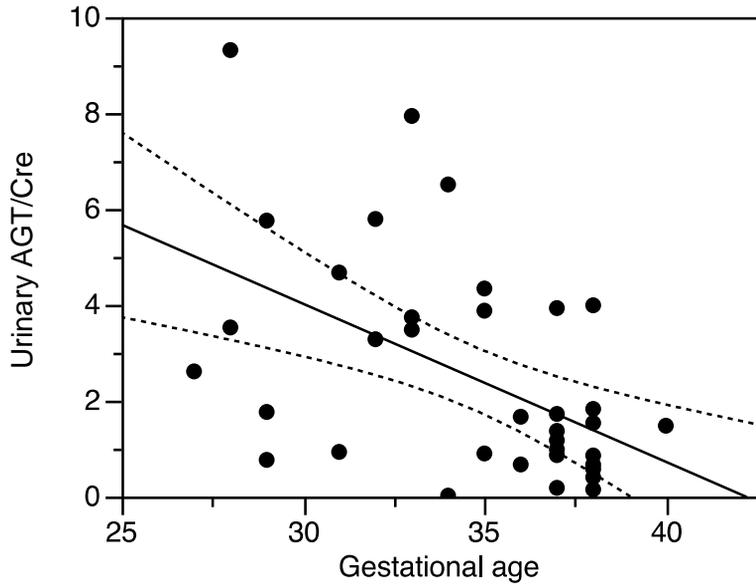


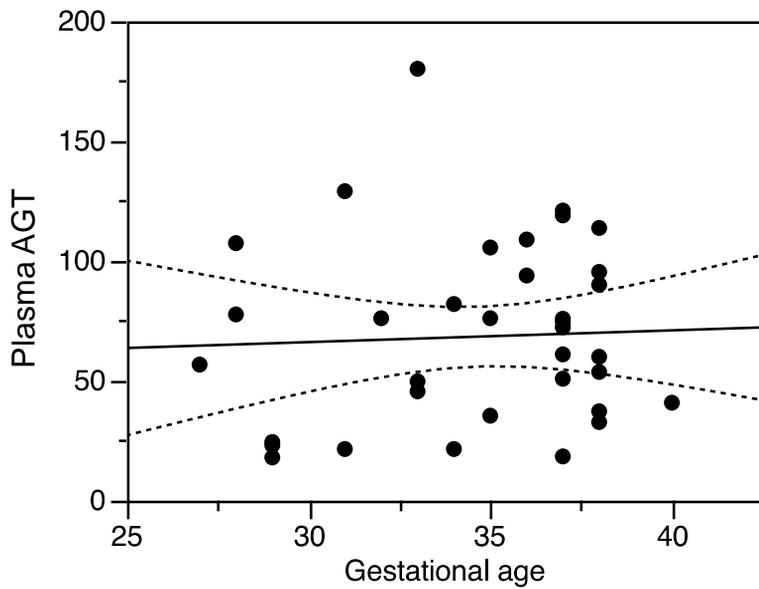
Figure 1

A



$$\text{Urinary AGT/Cre} = 13.91 - 0.33 \text{ Gestational age}$$
$$r = 0.5115, P = 0.0010$$

B



$$\text{Plasma AGT} = 51.54 + 0.49 \text{ Gestational age}$$
$$r = 0.0465, P = 0.7814$$

Figure 2

Table 1. Subject profiles

Parameters	Preterm N = 20	Full-term N = 18	P values	χ^2
Gestational age, weeks	32.0 +/- 0.6 **	37.7 +/- 0.2	< 0.01	
Birth weight, g	1,757 +/- 120 **	2,611 +/- 131	< 0.01	
Caesarean/vaginal delivery	10/10	8/10	0.73	0.12
Gender, F/M	9/11	7/11	0.70	0.15
Apgar score, 1 min	6.8 +/- 0.6 *	8.3 +/- 0.2	0.03	
Apgar score, 5 min	9.0 +/- 0.3	9.5 +/- 0.1	0.12	
Days of sampling after birth	2.5 +/- 0.3	2.0 +/- 0.3	0.25	
Plasma Cystatin C (mg/mL)	1.64 +/- 0.10	1.66 +/- 0.11	0.88	

F; Females, M; Males, *; $P < 0.05$, **; $P < 0.01$ vs. full-term.