

ORIGINAL**Sp6 downregulation of follistatin gene expression in ameloblasts**

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Abstract : Sp6 is a member of the Sp family of transcription factors that regulate a wide range of cellular functions, such as cell growth and differentiation. Sp6, also called epiprofin, is specifically expressed in tooth germ, limb bud, and hair follicle, but there is little information on its function.

To investigate the possible role of Sp6 in tooth development, first we established an Sp6-overproducing clone, CHA9, and analyzed the features of the cell, including cell proliferation and gene expression. The parental cells of CHA9 are the ameloblast-lineage G5 cells that we previously established from rat dental epithelia of lower incisor. Sp6 overproduction accelerated cell proliferation and induced the expression of ameloblastin mRNA, a marker of ameloblast differentiation. Second, we performed genome-wide screening of Sp6 target genes by microarray analysis. Out of a total 20,450 genes, 448 genes were up-regulated and 500 genes were down-regulated by Sp6. We found the expression of follistatin, a BMP antagonist, to be 22.4-fold lower in CHA9 than in control cells. Transfection of the Sp6-antisense construct into CHA9 cells restored follistatin expression back to equivalent levels seen in control cells, indicating that Sp6 regulates follistatin gene expression in ameloblasts.

Our findings demonstrate that the follistatin gene is one of the Sp6 target genes in ameloblasts and suggest that Sp6 promotes amelogenesis through inhibition of follistatin gene expression. *J. Med. Invest.* 55 : 87-98, February, 2008

Keywords : ameloblast, amelogenesis, follistatin, microarray, Sp

INTRODUCTION

Sp6, one of the Sp family members, is a tissue-specific transcription factor. Sp6 was previously called KLF14 (1). However, authentic KLF14 gene is localized at rat chromosome 2 and has been identified to be different from Sp6 gene (2). Epiprofin is another name of Sp6 based on its gene expres-

sion pattern and function (3). *In situ* hybridization analysis has revealed Sp6 mRNA expression in tooth germ, limb bud, and hair follicle (3). In newborn mouse incisors, Sp6 mRNA is expressed in early stages of tooth development to the secretory stage of ameloblasts. Sp6 mRNA is also weakly expressed in mesenchymal odontoblasts of the incisor (3). Recently, it has been reported that Sp6 mRNA is strongly expressed in embryos compared with adult tissues, suggesting a functional role in development and tissue homeostasis (4). However, there is no information about the levels of Sp6 protein expression in various tissues in the embryo.

Sp6 has a unique domain structure, as shown

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in Fig. 1A. It has three zinc finger motifs at the C-terminal region and two potential nuclear localization signals near the zinc finger domain. It is believed to bind to GC- or GT-rich DNA sequences, sharing common binding characteristics of all Sp and KLF family proteins (5). A proline-rich domain, presumed to be a transcriptional regulatory domain, at the N-terminal region, is also a unique characteristic of Sp5, Sp6, and Sp7 among the Sp family members (5). This proline-rich domain may provide unique functions in tooth development distinct from other Sp proteins (3). However, there is no information about the functional roles of Sp6 in transcriptional regulation involving target genes and interactive partners.

We explored the physiological roles of Sp6 in the developing tooth. We first prepared an antibody against rat Sp6 protein and confirmed the protein localization in the rat incisor. We also developed an Sp6-overproducing clone, CHA9. Next, we screened potential Sp6 target genes using microarray analysis by comparing expression in CHA9 with a control clone, V1. Among many potential target genes, we found significant downregulation of follistatin gene, which is an extracellular glycoprotein that directly binds to bone morphogenic protein (BMP) and acts by antagonizing BMP signals (6). Transfection of Sp6-antisense expression plasmid into CHA9 cells restored follistatin gene expression to the same level as control. It has been reported that follistatin is responsible for the formation of an enamel-free area in the mouse incisor and molar by inhibiting ameloblast differentiation (7-10).

Taken together, our findings demonstrate that Sp6 has a role in amelogenesis by inducing ameloblastin expression and inhibiting follistatin expression.

MATERIALS AND METHODS

Generation of anti-Sp6 antibody

Carboxyl-terminal peptides of rat Sp6, CGKGGK-REAEGSSASSN (XP_343962; amino acid 362-376), were conjugated with keyhole limpet hemocyanin (KLH) and used for immunization in rabbits (Fig. 1A) (Hokkaido System Sciences, Hokkaido, Japan). The specificity of antisera was analyzed by Western blotting of protein extracts of COS7 cells transfected with pCI-neo mammalian expression vector (Promega, Madison, WI) carrying the DNA fragment encoding FLAG-tagged rat Sp6 (Fig. 1B).

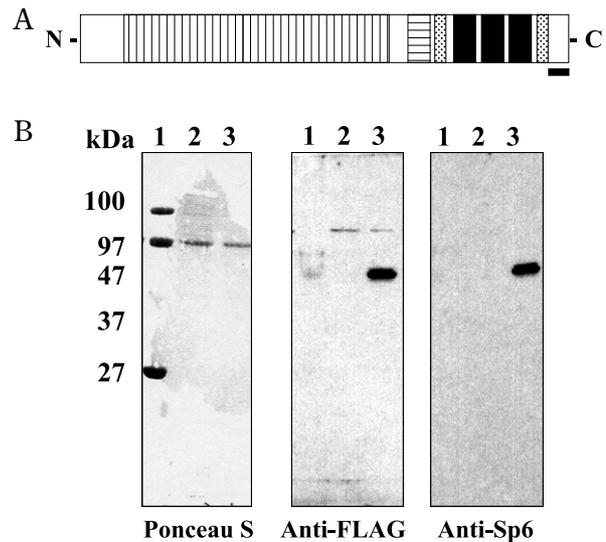


Fig. 1 Generation of rabbit polyclonal antibody against rat Sp6. (A) Structure of Sp6. Sp6 has several potential regulatory domains: a proline-rich region (vertical stripes), three zinc-finger domains (black boxes), and a Btd domain (horizontal stripes). Two candidate sequences for nuclear localization signals are also indicated (dotted pattern). The black bar indicates the position of the antigen epitope used to generate anti-Sp6 antibody. (B) Specificity of anti-Sp6 antibody. Western blot analysis was performed with protein extracts of COS7 cells transfected with pCI-neo mammalian expression vector carrying the DNA fragment encoding FLAG-tagged rat Sp6. Lane 1, molecular size marker; lane 2, COS7 cells transfected with empty vector; lane 3, COS7 cells transfected with FLAG-tagged rat Sp6 in pCI-neo.

Immunohistochemistry and cytochemistry

Lower mandibles were dissected from postnatal day 1 stroke-prone spontaneous hypertensive rats (SHRSP) and fixed with 4% paraformaldehyde (PFA). Four-micrometer thick sections were deparaffinized, rehydrated, and treated with 3% H₂O₂ in methanol to inhibit endogenous peroxidase activity. Antigens were retrieved by heat treatment using antigen unmasking solution (Vector Laboratories, Burlingame, CA). The sections were blocked with 3% horse serum in phosphate-buffered saline (PBS) without calcium and magnesium [PBS(-): 137 mM NaCl, 8.1 mM Na₂HPO₄ · 12H₂O, 2.68 mM KCl, 1.47 mM KH₂PO₄, pH 7.4] for 30 min at room temperature. Sections were incubated with our rabbit polyclonal anti-Sp6 antibody (1 : 2,000) or rabbit polyclonal anti-Amelogenin antibody (1 : 1,000; Hokkaido, Hokkaido, Japan) at 4°C overnight. After washing with PBS(-) for several times, sections were incubated with horseradish peroxidase-conjugated anti-rabbit IgG antibody (Histofine; Nichirei, Tokyo, Japan) for 30 min at room temperature. 3-3'-diaminobenzidine (DAB) solution was used to detect immune-complex ac-

cording to the manufacturer's instructions (Nihirei). The sections were then counterstained with hematoxylin. The stained sections were observed under a light microscope (BX51-34 FL-1-K-O ; Olympus, Tokyo, Japan).

COS7 cells were transfected with pCI-neo mammalian expression vector (Promega, Madison, WI) carrying the DNA fragment encoding FLAG-tagged rat Sp6 or vector alone as a negative control, using Lipofectamine plus reagent (Invitrogen, Carlsbad, CA) according to manufacturer's instructions. Twenty-four hours after transfection, the cells were rinsed with PBS(-), fixed with ethanol for 10 min at room temperature, permeabilized by 0.1% Triton X-100 in PBS(-), and blocked with 4% BSA/PBS(-) for 30 min at room temperature. The cells were then incubated with mouse anti-FLAG M2 monoclonal antibody (1 : 500 ; 2 mg/ml ; Sigma, St. Louis, MO) for 1 hr at room temperature followed by incubation with goat anti-mouse IgG (H + L) conjugated with Alexa Fluor-488 (Invitrogen). The cells were stained with 2 mg/ml of Hoechst 33342 (Sigma), and mounted in Vectashield mounting medium (Vector Laboratories, Burlingame, CA). The samples were observed under a fluorescence microscope (Olympus BX51 ; Olympus, Tokyo, Japan). The images of both immunohistochemistry and cytochemistry were captured by a digital imaging system (DP-70 set ; Olympus).

Establishment of Sp6-stable transformants

An Sp6 expression vector was constructed as follows. The C'- or N'-terminals of the Sp6 coding region were tagged with HA and ligated into pCIneo expression vector (Sp6-CHA or Sp6-NHA in pCIneo). Full-length coding regions of Sp6 fused with HA-tag and *SpeI* site at 3'-end were generated by PCR. The PCR product was cloned into pGEM-T easy vector (Promega) and confirmed by restriction enzyme digestion and sequencing. The plasmid was named Sp6-CHA in pGEM-T easy. To construct an expression vector, Sp6-CHA fragment was produced by *NotI* digestion from Sp6-CHA in pGEM-T easy and re-cloned into pCI-neo vector (Promega). The final construct (Sp6-CHA in pCIneo) was confirmed by restriction enzyme digestion and sequencing.

Sp6-stable transformants were established using an ameloblast-lineage clone, G5, as described previously (11). G5 cells were plated in 100 mm culture dishes and maintained in Dulbecco's modified Eagle's medium (DMEM)/Ham's F12 medium (Nissui, Tokyo, Japan) with 10% fetal bovine serum

(FBS ; JRH Biosciences, Lenexa, KS). Sp6-CHA, Sp6-NHA, and empty vector were stably transfected into G5 cells using Lipofectamine plus reagent as described in the section of Immunohistochemistry and cytochemistry in MATERIALS AND METHODS. For long-term growth, the cells were maintained with selection medium containing 1 mg/ml of G418. G418-resistant clones were isolated by limiting dilution.

Cell proliferation assay

To analyze cell growth activity, we performed an Alamar Blue assay (BioSource, Camarillo, CA) as described previously (12). In brief, the cells were inoculated in 96-well dishes ($\sim 10^3$ cells/well), the medium was changed to fresh DMEM/Ham's F12 without FBS after 24 h. Alamar blue reagent was added to each well to attain a final concentration of 10% (v/v). Cells were cultured for further 4 h. The absorbance of each sample was measured at 570/595 nm using a Multiskan JX microplate reader (Thermo Lab systems, Tokyo, Japan).

Western blot analysis

To identify an Sp6-overproducing clone, Western blot analysis was performed. Stably transformed cells were dissolved in 2X sodium dodecyl sulfate (SDS) sample buffer (125 mM Tris-HCl pH 6.8, 4% SDS, 10% glycerol). Protein concentration was determined using the BCA Protein Assay Kit (Pierce, Rockford, IL). After adding 10% 2-mercaptoethanol and 0.01% bromophenol blue, samples were boiled for 3 min at 95°C. Eighty micrograms of protein was resolved on 12.5% SDS-polyacrylamide gels. After electrophoresis, the proteins were transferred to a polyvinylidene difluoride (PVDF) membrane (Immobilon™-P ; Millipore, Bedford, MA) and blocked with 5% (w/v) skim milk in Tris-buffered saline and Tween-20 (TBS-T ; 10 mM Tris-HCl, 150 mM NaCl, 0.05% (v/v) Tween-20, pH 8.0) at 4°C overnight. The membrane was incubated with anti-HA-probe Y11 (1 : 1,000 ; sc-805 ; Santa Cruz Biotechnology, Santa Cruz, CA) in the same buffer as that used for blocking, at room temperature for 1 h. After washing several times in TBS-T, the membrane was incubated with horseradish peroxidase-conjugated anti-rabbit IgG antibody (1 : 5,000 ; Amersham Biosciences, Buckinghamshire, UK) for 30 min at room temperature. After washing several times, the immune complex was detected using the ECL Plus western blotting detection reagents (Amersham Biosciences) according to the manufacturer's instructions.

Microarray analysis

Both V1 and CHA9 cells were harvested and total RNA was isolated using the RNeasy[®] mini kit (Qiagen Inc., Valencia, CA). cDNA was synthesized from 500 ng of total RNA using M-MLV Reverse Transcriptase (Invitrogen) and was labeled with Cy3 and Cy5, respectively. The labeled probes were hybridized onto a rat oligo DNA microarray containing 22,450-oligo DNAs on the slide (Agilent Technologies, Santa Clara, CA). The arrays were scanned, and the intensity and ratio of the two fluorescent signals of Cy3 and Cy5 were analyzed by Agilent Feature Extraction software (Agilent). The raw data were unified to calibrate the systemic error between the Cy5 and Cy3 labeling steps using the expression levels of the housekeeping genes spotted on the array. Differential expression of the genes was determined by the following criteria: gene expression in CHA9 was considered to be up-

regulated if the intensity value was increased two folds compared to V1 (Cy5/Cy3 > 2). Conversely, gene expression was considered down-regulated when the ratio of Cy5/Cy3 signals is less than 0.5.

RT-PCR analysis

To confirm the microarray results, we selected several up-regulated and down-regulated genes that are related to the differentiation markers or signaling molecules including Crp2, a LIM-only protein (13); Dec2, a member of the basic helix-loop-helix superfamily (14); Cyp26b, a p450 isoform (15); follistatin, a BMP antagonist (7); and fibronectin, an extracellular matrix (16), and performed RT-PCR analysis with gene-specific primers. Five hundred nanograms of total RNA was used for reversed transcription with an RNA PCR kit (Takara, Shiga, Japan). The synthesized cDNA was used as a template for PCR. Gene-specific primers and PCR conditions are shown in Table 1.

Table 1 Gene-specific primers.

Gene	Primer	Annealing (°C)	Extension (sec)	Cycles
GAPDH	5'-CATTGACCTCAACTACATGG-3' 5'-CTCAGTGTAGCCCAGGATGC-3'	58	60	20
Fibronectin	5'-GAAAGGCAACCAGCAGAGTC-3' 5'-CTGGAGTCAAGCCAGACAC-3'	58	30	30
Follistatin	5'-TTTTCTGTCCAGGCAGCTCCAC-3' 5'-GCAAGATCCGGAGTGCTTCACT-3'	58	30	25
Cyp26b	5'-AGGGCCTAGAGAAGGCTGTC-3' 5'-GTTCTCCCGTAGCTTTTCC-3'	58	30	30
Jagged1	5'-GTCCACGGCACCTGCAATG-3' 5'-ACAAGGCTTGGCCTCGCAC-3'	58	60	25
Dec2	5'-AACATCCCCTAAGGCGTACC-3' 5'-CTCTCCCACCCAACAGACTC-3'	58	30	30
Crp2	5'-TGTACCACGCTGAAGAGGTG-3' 5'-CCCAAAGTTCTTTGCGTAGC-3'	58	30	30
Ameloblastin	5'-CATGTCTTATGGAGCAAACCA-3' 5'-TCTTGCAAGTGGAGAGCCTTCT-3'	60	30	40
Amelogenin	5'-CAAGAAATGGGGACCTGGATC-3' 5'-GCTGCCTTATCATGCTCTGG-3'	60	30	40
Sp1	5'-TCAGCTGCCACCATGAGCG-3' 5'-TGGGTGACTCAATTCTGCTGC-3'	58	30	30
Sp2	5'-TCAGTCCCAGTACTACTG-3' 5'-CCTGGATGGTCTGGGAACG-3'	58	30	30
Sp3	5'-ACTTTGACGCCTGTTTCAGAC-3' 5'-TGATGTTGTTGGTCCCCTTC-3'	58	30	30
Sp4	5'-ACACCTTCAGGGCAAATCAG-3' 5'-GCTTGCTGGACTTTCACTCC-3'	58	30	30
Sp5	5'-TCCTTCAGGTGCCCTATGAC-3' 5'-GGTAGCAGGTTGGAGTAGCC-3'	58	45	35
Sp6	5'-CCGGCAATGCTAACCGCTGTCTGTG-3' 5'-GGCTCAGTTGGAGGACGCCGAGCTG-3'	58	30	30
Sp7	5'-CACTGGCTCCTGGTTCTCTC-3' 5'-AGGAAATGAGTGGGGAAAGG-3'	58	30	30
Sp8	5'-CTCTCTAGCTTCGGCGTGTG-3' 5'-TGACCTTGAGATGAACAC-3'	58	45	35
Sp9	5'-GCCTTCATCTCCAAGGTGC-3' 5'-GTGGAGCTAAAAGCCGAGTG-3'	58	45	35

Transient transfection with Sp6-antisense construct

Sp6-antisense expression plasmid was constructed by inversely inserting the rat Sp6 coding DNA fragment into pCI neo vector. The Sp6-antisense construct was transiently transfected into CHA9 cells in 6-well plates, using Lipofectamine plus reagent (Invitrogen) as described in the section of Immunohistochemistry and cytochemistry in MATERIALS AND METHODS. After 24 h, at about 80% confluence, V1 or CHA9 cells were transfected with 1 μ g of Sp6-antisense expression plasmid. Forty-eight hours later, cells were harvested and levels of *follicle-statin* gene expression were determined by RT-PCR.

RESULTS

Characterization of anti-Sp6 antibody

We first developed a rabbit polyclonal antibody against the C-terminal region of rat Sp6 epitope. To check the reactivity and specificity of this antibody, COS7 cells were transiently transfected with FLAG-tagged Sp6 or empty vector as a negative control. Cell lysates were electrophoresed and analyzed by immunoblotting. Both anti-Sp6 and anti-FLAG antibodies detected a discrete single band at the same molecular mass, 47 kDa, which was not detectable in the control, indicating that the antibody is specific to Sp6 without background (Fig. 1B).

Localization of Sp6 in vivo

To examine the expression pattern of Sp6 in the tooth, we performed immunohistochemical analysis using rat lower incisors at postnatal day 6, because all developmental stages can easily be seen. As shown in Fig. 2A, Sp6 was broadly detected in the ameloblast layer (Fig. 2A, upper). Amelogenin, one of the enamel proteins and a differentiation marker of amelogenesis, was also detected in the same location as that of Sp6 (Fig. 2A, middle). At high magnification, Sp6 was clearly detectable mainly in the nucleus of differentiating ameloblasts, which are tall, columnar, and polarized cells, and in some odontoblasts (Fig. 2B). Interestingly, Sp6 was not detected in all ameloblasts at the same differentiating stages.

The intracellular localization of Sp6 was also confirmed in Sp6-transfected COS7 cells by immunocytochemistry with anti-FLAG M2 monoclonal antibody (Fig. 3). Fluorescent signals were detected

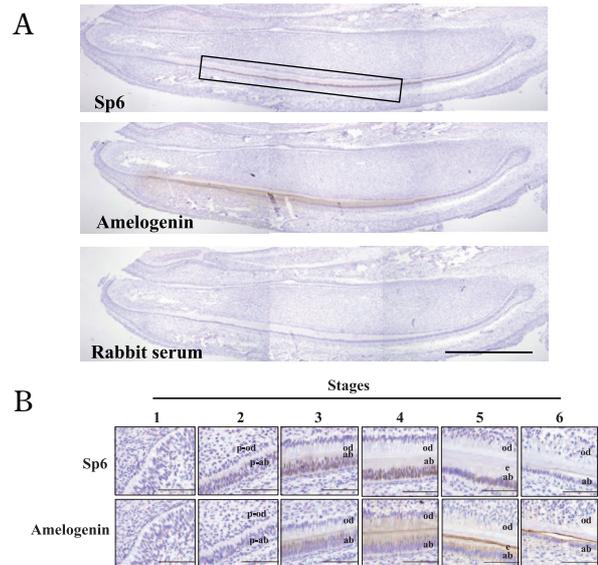


Fig. 2 Localization of Sp6 protein in rat incisor.

(A) Overview of Sp6 localization in incisor of 6-day-old rat mandible. Amelogenin and Sp6 were mainly detected in ameloblasts. Scale bar indicates 1.0 mm. The box indicated the area of position of Sp6-positive cells. (B) The stage-specific localization of Sp6 in amelogenesis. Stage 1, undifferentiated; stage 2, pre-ameloblast; stage 3, pre-secretory; stage 4, secretory; stage 5, maturation; stage 6, reduced. From the pre-ameloblast to the maturation stages, Sp6 was detected mainly in the nucleus and weakly in cytosol. Sp6 was also detected in some odontoblasts. enamel; e, odontoblasts; od, ameloblasts; ab, preodontoblasts; p-ab, preameloblasts; p-ab. Scale bar indicates 100 μ m.

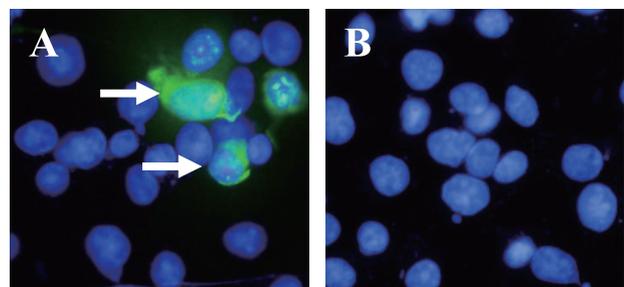


Fig. 3 Subcellular localization of Sp6.

COS7 cells were transfected with pCI-neo vector carrying FLAG-tagged SP6 (A), or mock vector as a negative control (B). Sp6 was detected by anti-FLAG antibody and Alexa 488-conjugated secondary antibody. Green color indicates Sp6 localization and blue color indicates nucleus stained with Hoechst 33342.

only in Sp6-transfected samples (Fig. 3A), whereas no fluorescence from the secondary antibody was detected in mock-transfected control with vector alone (Fig. 3B). The signals were detected in both cytoplasm and nucleus. The dotted-like fluorescence signals were often observed in the nucleus (Fig. 3A).

Establishment of an Sp6-overproducing clone, CHA9

We prepared several HA-tagged Sp6 expression plasmids, and stably transformed an ameloblast-

lineage cell line, G5 (11). More than 100 clones were selected by G418 and established. The level of Sp6 production was screened by immunoblot analysis and the highest Sp6 producer, CHA9, was selected (Fig. 4, Lane 4). The effects of Sp6 overproduction on cell growth were investigated in some representative clones. CHA9 cells had the most accelerated growth rate compared with other clones as shown in Fig. 5. We next examined the effects of Sp6 overproduction on the gene expression of other Sp family members, since they are involved in growth regulation, differentiation, signal transduction, and morphogenesis (3, 17-25). The expression

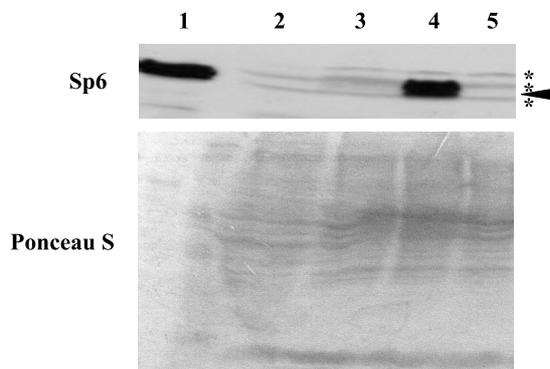


Fig. 4 Screening for Sp6 high producer clone. (A) Lane 1, Sp6-CHA in pCIneo transfected into COS7 cells as a positive control ; lane 2, empty vector transfected into G5 cells as a negative control ; lane 3, Sp6-CHA in pCI-neo transfected clone 2 (CHA2) ; lane 4, Sp6-CHA in pCI-neo transfected clone 9 (CHA9) ; lane 5, Sp6-NHA in pCI-neo transfected clone 16 (NHA16). A representative result is shown. Ponceau S staining was performed as a loading control. Asterisk ; non-specific band, arrowhead ; Sp6 protein.

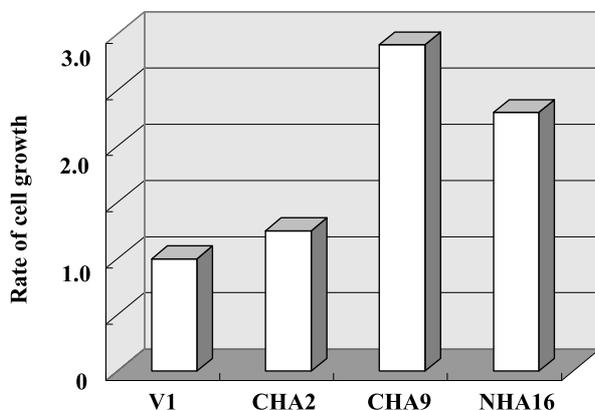


Fig. 5 Effects of Sp6 overproduction on cell growth. The rate of cell growth was determined by Alamar Blue analysis. Vertical axis indicates the relative rate of cell growth compared with empty vector-transfected G5 (V1). CHA2, CHA9, and NHA16 corresponded to the transformants that were transfected with different tagged Sp6 expression constructs. The highest Sp6-producer clone, CHA9, had the most accelerated cell growth.

levels of other Sp family genes were not affected in CHA9 cells, except for Sp8 that was weakly induced (Fig. 6A). Sp5, Sp7, and Sp9 were not expressed in CHA9 and V1 cells (Fig. 6A). Levels of mRNA of ameloblastin and amelogenin, two major enamel proteins that are ameloblast differentiation markers, were also assessed by RT-PCR analysis (Fig. 6B). The level of ameloblastin mRNA was significantly increased in CHA9 cells, although that of amelogenin was not changed.

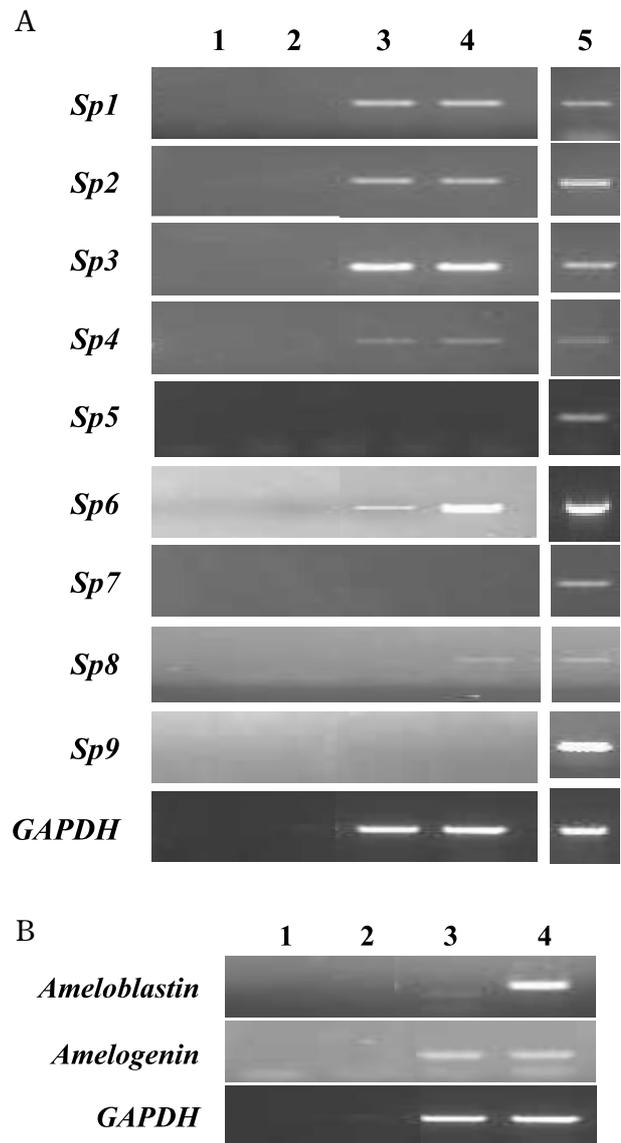


Fig. 6 Gene expression patterns of Sp family members and enamel proteins.

(A) Expression levels of Sp family members determined by RT-PCR. Lane 1, negative control (template ; water) ; lane 2, negative control (template ; no reverse-transcriptase, RT(-)) ; lane 3, control (template ;V1), lane 4, Sp6 overproducer (template ; CHA9) ; lane 5, positive control for PCR (template ; rat day 6 molar).

(B) Expression levels of amelogenin and ameloblastin determined by RT-PCR on the same samples used in (A).

Identification of possible Sp6 target genes by microarray analysis

To identify the possible target genes of Sp6, we screened a total of 20,450 genes by microarray analysis. RNA samples were prepared from the Sp6-overproducing clone, CHA9, and the control clone, V1, and hybridized onto the specific oligonucleotides to each gene in the microarray. Differentially expressed genes were selected on the basis of a greater than 2-fold or less than 0.5-fold difference in the expression levels between CHA9 and V1 cells. We found that 448 genes were up-regulated and 500 genes were down-regulated by Sp6 overproduction in ameloblast-lineage clone, and these are summarized in Table 2. The regulated genes were

Table 2 Summary of Sp6 gene regulation by microarray analysis.

	Known	Unknown	Total
Up	306	142	448
Down	382	118	500
Total	688	260	948

also classified by major cellular functions, as shown in Table 3. Both top 30 up-regulated and down-regulated genes are listed in Table 4 and 5, respectively.

Table 3 Functional classification of Sp6 potential target genes.

Category	Up-regulated	Down-regulated
Apoptosis	6	6
Cell adhesion	5	9
Cell growth	4	6
Channel	16	11
Cytoskeleton	2	5
Enzyme	87	101
ECM	2	5
Extracellular signaling molecule	3	20
Transcriptional regulation	15	23
Peptidolysis	0	1
Receptor	30	22
Signal transduction	22	26
Other	114	147
Total	306	382

Table 4 Up-regulated genes in CHA9.

#	up-regulated genes	C9/V1 (fold)
1	Q 60501 Hamster p7 preinsertion DNA [TC540005]	25.1
2	Rat alpha-methylacyl-CoA racemase, mRNA [NM_012816]	24.9
3	Rat endothelin receptor type B, mRNA [NM_017333]	19.5
4	Rat gap junction membrane channel protein bet, mRNA [NM_001004099]	18.6
5	Rat glutamate receptor interacting protein 1, mRNA [NM_032069]	17.5
6	Rat collagen, type V, alpha 3, mRNA [NM_021760]	16.8
7	Rat lectin, galactose binding, soluble 7, mRNA [NM_022582]	16.3
8	Rat Dock1 protein, mRNA [XM_219424]	16.3
9	Rat surfactant associated protein C, mRNA [NM_017342]	13.2
10	Q8R4C3 Csr1 [TC 556501]	11.3
11	Rat liver glycogen phosphorylase, mRNA [NM_022268]	11.2
12	MOUSE P2Y purinoceptor 5 [TC521815]	10.8
13	Rat serine/threonine kinase 39, STE20/SPS1 homolog, mRNA [NM_019362]	10.5
14	Rat nuclear receptor subfamily 3, group C, member 2, mRNA [NM_013131]	10.4
15	Rat brain derived neurotrophic factor, mRNA [NM_012513]	10.0
16	Rat glutathione-S-transferase, alpha type2, mRNA [NM_017013]	10.0
17	Rat solute carrier family 1, member 3, mRNA [NM_019225]	9.2
18	Rat glutathione S-transferase, alpha 4, mRNA [XM_217195]	9.0
19	MOUSE Asporin [TC557668]	9.0
20	Rat calcium channel, voltage-dependent, alpha2/delta subunit 1, mRNA [NM_012919]	8.6
21	Rat cytosolic cysteine dioxygenase 1, mRNA [NM_052809]	8.6
22	Rat sialyltransferase 10 (alpha-2,3-sialyltransferase VI), mRNA [NM_207602]	8.5
23	Rat basic helix-loop-helix domain containing, class B3 (Dec2), mRNA [NM_133303]	8.4
24	Q70US4 NADH dehydrogenase 6 [TC525243]	8.4
25	Rat transmembrane 7 superfamily member 3, mRNA [NM_001011970]	8.2
26	Rat leukotriene B4 12-hydroxydehydrogenase, mRNA [NM_138863]	7.7
27	Rat desmoglein 5, mRNA [XM_214616]	7.4
28	Rat carbohydrate (keratan sulfate Gal-6) sulfotransferase 1, mRNA [NM_001011955]	7.4
29	Rat glutamate cysteine ligase, modifier subunit, mRNA [NM_017305]	7.0
30	Rat NEDD9 interacting protein with calponin homology and LIM domains, mRNA [XM_215424]	6.8

Table 5 Down-regulated genes in CHA9.

#	down-regulated genes	V1/C9 (fold)
1	Rat interleukin 1 receptor-like 1, mRNA [NM_013037]	144.2
2	Rat S100 calcium binding protein A9, mRNA [NM_053587]	61.0
3	Rat complement component 3, mRNA [NM_016994]	58.8
4	Rat matrix metalloproteinase 10, mRNA [NM_133514]	51.7
5	Ratsolute carrier family 28, member 2, mRNA [NM_031664]	37.0
6	Mouse Derlin-3 [TC555644]	30.6
7	Rat schlafen 3, mRNA [NM_053687]	26.5
8	Rat interleukin 1 alpha, mRNA [NM_017019]	25.2
9	Rat follistatin, mRNA [NM_012561]	22.4
10	Rat latent transforming growth factor beta binding protein 1, mRNA [NM_021587]	22.2
11	Q8HXG3 P76RBE protein [TC539175]	20.5
12	Rat colony stimulating factor 3, mRNA [NM_017104]	20.5
13	Rat decay accelerating factor 1, mRNA [BC061869]	19.5
14	Rat kidney androgen regulated protein, mRNA [NM_052802]	18.0
15	Rat H 2.0-like homeo box 1, mRNA [XM_344184]	17.8
16	Rat keratin complex 2, basic, gene 7, mRNA [XM_217035]	17.3
17	Rat hydroxysteroid 11-beta dehydrogenase 1, mRNA [NM_017080]	15.6
18	Rat chemokine (C-X-C motif) ligand 5, mRNA [NM_022214]	15.6
19	Rat 15-hydroxyprostaglandin dehydrogenase, mRNA [NM_024390]	15.2
20	Rat matrix metalloproteinase 3, mRNA [NM_133523]	14.7
21	Rat chemokine (C-X-C motif) ligand 2, mRNA [NM_053647]	14.6
22	Rat transglutaminase 2, C polypeptide, mRNA [NM_019386]	14.1
23	Rat secretory leukocyte peptidase inhibitor, mRNA [NM_053372]	13.6
24	Rat CRIB-containing BORG3 protein, mRNA [XM_341784]	13.3
25	Rat serine protease, mRNA [XM_220222]	13.2
26	Rat mRNA for granzyme-like protein III. [X 76996]	12.7
27	Rat coagulation factor 3 mRNA [NM_013057]	12.4
28	Rat regulator of G-protein signaling 2, mRNA [NM_053453]	12.4
29	Rat ischemia related factor vof-16, mRNA [NM_147207]	12.3
30	Rat protein tyrosine phosphatase, non-receptor type 5, mRNA [NM_019253]	12.3

Validation of Sp6-regulated genes by RT-PCR

RT-PCR analysis showed that expression levels of Jagged 1, Crp2, and Dec2 were enhanced in CHA9 cells compared with V1 (Fig. 7A), whereas those of Cyp 26b, follistatin, and fibronectin were reduced (Fig. 7B). Especially, the expression of follistatin was greatly down-regulated by Sp6 overexpression.

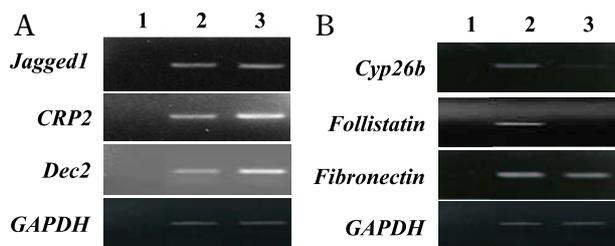


Fig. 7 Confirmation of microarray results by RT-PCR. (A) Representative up-regulated genes in CHA9. RT-PCR was performed on the same samples as in Fig. 6. Lane 1, negative control (RT(-)); lane 2, V1; lane 3, CHA9. (B) Representative down-regulated genes in CHA9. Lane 1, negative control (RT(-)); lane 2, V1; lane 3, CHA9.

Follistatin gene expression restored by Sp6-antisense construct

In order to establish whether Sp6 directly regulates follistatin gene expression, a loss-of-function experiment was performed by transfecting an Sp6-antisense expression plasmid into CHA9 and V1 cells. After transfection, total RNA was isolated and the follistatin gene expression was analyzed by RT-PCR. The results showed that Sp6 knockdown restored follistatin gene expression to almost the same level as that in controls (Fig. 8A and B). We also analyzed whether ameloblastin gene expression was down-regulated by Sp6 knockdown technique. The level of ameloblastin mRNA in Sp6-antisense treated CHA9 cells was 76 % reduced compared to that of no treated CHA9 cells (data not shown). The levels of Sp6 protein were reciprocally correlated with follistatin gene expression (Fig. 8C). In V1 cells, on the other hand, transfection with Sp6-antisense expression plasmid did not cause any significant changes to follistatin expression levels. Sp6 protein

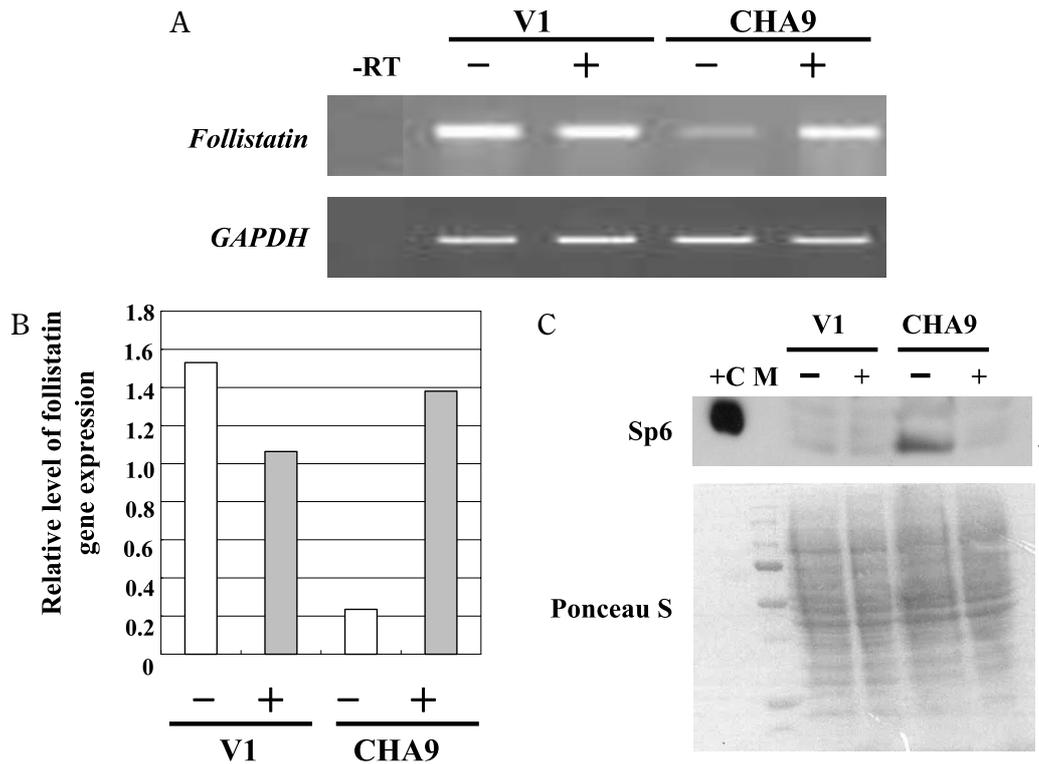


Fig. 8 Restoration of follistatin gene expression by Sp6 knockdown in CHA9 cells. Sp6-antisense construct was transiently transfected (+) or not (-) into V1 and CHA9 cells, respectively. (A) RT-PCR results of follistatin gene expression. (B) Relative ratio of follistatin gene expression. RT-PCR products in (A) were measured by Quantity One (BIO-RAD). (C) The effect of Sp6-knockdown on Sp6 protein level. Upper panel shows western blot analysis. Lower panel shows Ponceau S staining pattern as a loading control. Sp6 protein was detected by anti-Sp6 antibody (arrowhead). +C ; a positive control (same as lane 1 in Fig.4), M ; molecular size marker.

was not detected in V1 cells although Sp6 mRNA was weakly detected as shown in Fig. 6A.

DISCUSSION

Sp6 is widely believed to be a DNA-binding transcription factor that may have diverse roles in tooth development and morphogenesis (3, 5). Sp6 mRNA was found to be expressed in a tissue-specific manner in tooth germ, hair follicle, and limb bud (3). However, there is no evidence that Sp6 is a transcription factor.

In this study, we first developed an antibody against Sp6 protein (Fig. 1). By immunohistochemical analysis, we detected Sp6 expression mainly in the nucleus of ameloblasts in the presecretory and secretory stages, and in some odontoblasts (Fig. 2B). Interestingly, we found that amelogenin expression was localized to the same region as that of Sp6 expression (Fig. 2A, middle). Next, we investigated Sp6 target genes with the combined methods of gain of function analysis and microarray analysis. We established an Sp6-overproducing clone, CHA9,

which was transformed by Sp6 expression plasmid using ameloblast-lineage clone, G5 (11). CHA9 cells grow relatively much faster than control cells, which is consistent with previous reports on the cloning of Sp6 (3). Analysis for the effects of Sp6 overproduction on the gene expression of other Sp family members revealed no gross difference in CHA9 cells compared with control, V1 cells, except for that of Sp8 that was slightly induced in CHA9 cells (Fig. 6A). The physiological role of Sp8 in ameloblasts remains to be determined, although it has been reported that Sp8 is expressed during brain and limb bud development and demonstrated to regulate cortical patterning and limb outgrowth through FGF8 induction (26, 27). In addition, gene expression of ameloblastin, one of the enamel-producing proteins, was significantly up-regulated in CHA9 cells compared with V1 cells (Fig. 6B), although the expression of amelogenin gene was not changed.

These findings suggested that Sp6 may have a regulatory role in tooth development and that Sp6 targets specific genes. We attempted to identify these target genes by pulling out differentially expressed genes by microarray analysis using RNA samples

from CHA9 and V1 cells. A total of 948 genes were found to differ in expression levels by more than 2-fold or less than 0.5-fold, and these were considered to be potential Sp6 target genes (Table 2). The genes could be divided into several functional categories, including transcriptional regulation, cell cycle/apoptosis, metabolism, cell adhesion, enzymatic, ion channel, cytoskeletal, extracellular matrix, extracellular signaling, proteolysis, receptor, and signal transduction (Table 3), suggesting many diverse roles for Sp6. Some of the up-regulated and down-regulated genes were related to tooth development. Among these, we focused on follistatin gene since it was greatly suppressed by Sp6 expression and also it has been reported that follistatin may inhibit ameloblast differentiation (7, 8). Follistatin is localized in enamel-free areas of mouse incisors, and follistatin transgenic mouse caused enamel defect in incisor (7, 8). Furthermore, follistatin knockout mouse showed no enamel free area of the incisors (7, 8). In this study, we found that ameloblastin gene expression was enhanced by Sp6 (Fig. 6B), and that Sp6 was co-localized with amelogenin in enamel-forming regions (Fig. 2), suggesting a functional involvement of Sp6 in enamel formation and amelogenesis. We also found that transfection of an Sp6-antisense construct restored follistatin gene expression in CHA9 cells almost to the levels of V1 cells, supporting a role of Sp6 in the regulation of follistatin gene expression.

Taken together, our findings suggest that Sp6 may regulate amelogenesis through the downregulation of follistatin gene expression. This is consistent with previous reports that demonstrated the involvement of follistatin regulation in BMP-mediated amelogenesis (7, 8). Follistatin is also known as an antagonist for activin. There are several reports to suggest a role of activin in tooth development (7, 9, 28-30). However, the expression of activin has been limited at the very early stage in the dental epithelial cells, and at the late stage in mesenchymal cells during tooth development (7, 9, 28-30). These findings suggested that follistatin may function in antagonizing BMPs in ameloblasts.

Figure 9 shows the interaction between Sp6, follistatin, and BMP in amelogenesis. We propose a role for Sp6 in amelogenesis through the downregulation of follistatin gene expression during tooth development and morphogenesis.

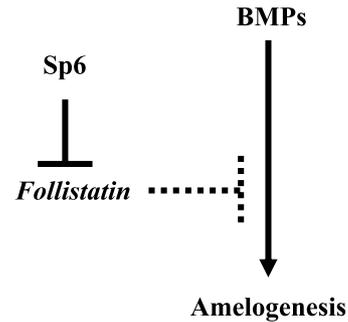


Fig. 9 Proposed model for the role of Sp6 in amelogenesis. Based on our gain-of-function and loss-of-function data, we propose a role for Sp6 in amelogenesis through down-regulation of follistatin gene expression. Sp6 downregulates follistatin gene expression, then it may promote amelogenesis.

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