

SCIENTIFIC REPORTS



OPEN

A genome-wide association study in the Japanese population identifies the 12q24 locus for habitual coffee consumption: The J-MICC Study

Hiroko Nakagawa-Senda^{1,2,10}, Tsuyoshi Hachiya³, Atsushi Shimizu³, Satoyo Hosono¹, Isao Oze⁴, Miki Watanabe¹, Keitaro Matsuo^{4,5}, Hidemi Ito^{4,5}, Megumi Hara⁶, Yuichiro Nishida⁶, Kaori Endoh⁷, Kiyonori Kuriki⁷, Sakurako Katsuura-Kamano⁸, Kokichi Arisawa⁸, Yora Nindita⁹, Rie Ibusuki⁹, Sadao Suzuki¹⁰, Akihiro Hosono¹⁰, Haruo Mikami¹¹, Yokko Nakamura¹¹, Naoyuki Takashima¹², Yasuyuki Nakamura¹³, Nagato Kuriyama¹⁴, Etsuko Ozaki¹⁴, Norihiro Furusyo¹⁵, Hiroaki Ikezaki¹⁵, Masahiro Nakatochi¹⁶, Tae Sasakabe², Sayo Kawai², Rieko Okada², Asahi Hishida², Mariko Naito², Kenji Wakai², Yukihide Momozawa¹⁷, Michiaki Kubo¹⁷ & Hideo Tanaka¹

Coffee is one of the most widely consumed beverages worldwide, and its role in human health has received much attention. Although genome-wide association studies (GWASs) have investigated genetic variants associated with coffee consumption in European populations, no such study has yet been conducted in an Asian population. Here, we conducted a GWAS to identify common genetic variations that affected coffee consumption in a Japanese population of 11,261 participants recruited as a part of the Japan Multi-Institutional Collaborative Cohort (J-MICC) study. Coffee consumption was collected using a self-administered questionnaire, and converted from categories to cups/day. In the discovery stage ($n = 6,312$), we found 2 independent loci (12q24.12–13 and 5q33.3) that met suggestive significance ($P < 1 \times 10^{-6}$). In the replication stage ($n = 4,949$), the lead variant for the 12q24.12–13 locus (rs2074356) was significantly associated with habitual coffee consumption ($P = 2.2 \times 10^{-6}$), whereas the lead variant for the 5q33.3 locus (rs1957553) was not ($P = 0.53$). A meta-analysis of the discovery and replication populations, and the combined analysis using all subjects,

¹Division of Epidemiology and Prevention, Aichi Cancer Center Research Institute, Nagoya, Japan. ²Department of Preventive Medicine, Nagoya University Graduate School of Medicine, Nagoya, Japan. ³Division of Biomedical Information Analysis, Iwate Tohoku Medical Megabank Organization, Disaster Reconstruction Center, Iwate Medical University, Morioka, Japan. ⁴Division of Molecular and Clinical Epidemiology, Aichi Cancer Center Research Institute, Nagoya, Japan. ⁵Department of Epidemiology, Nagoya University Graduate School of Medicine, Nagoya, Japan. ⁶Department of Preventive Medicine, Faculty of Medicine, Saga University, Saga, Japan. ⁷Laboratory of Public Health, Division of Nutritional Sciences, School of Food and Nutritional Sciences, University of Shizuoka, Shizuoka, Japan. ⁸Department of Preventive Medicine, Institute of Biomedical Sciences, Tokushima University Graduate School, Tokushima, Japan. ⁹Department of International Island and Community Medicine, Kagoshima University Graduate School of Medical and Dental Sciences, Kagoshima, Japan. ¹⁰Department of Public Health, Nagoya City University Graduate School of Medical Sciences, Nagoya, Japan. ¹¹Division of Cancer Prevention and Epidemiology, Chiba Cancer Center, Chiba, Japan. ¹²Department of Public Health, Shiga University of Medical Science, Shiga, Japan. ¹³Department of Food Science and Human Nutrition, Faculty of Agriculture, Ryukoku University, Kyoto, Japan. ¹⁴Department of Epidemiology for Community Health and Medicine, Kyoto Prefectural University of Medicine, Kyoto, Japan. ¹⁵Department of Environmental Medicine and Infectious Disease, Kyushu University, Fukuoka, Japan. ¹⁶Center for Advanced Medicine and Clinical Research, Nagoya University Hospital, Nagoya, Japan. ¹⁷Laboratory for Genotyping Development, Center for Integrative Medical Sciences, RIKEN, Kanagawa, Japan. Correspondence and requests for materials should be addressed to H.N.-S. (email: hirokosenda612@gmail.com)

	Discovery stage	Replication stage	All subjects (Discovery and Replication stage)
Number	6,312	4,949	11,261
Female (%)	55.0	52.6	53.9
Age \pm SD (year)	53.0 \pm 9.9	55.1 \pm 8.8	54.0 \pm 9.5
Coffee consumption (mean \pm SD) (cups/day)	1.6 \pm 1.5	1.7 \pm 1.6	1.7 \pm 1.5
Current alcohol drinkers (%)	56.1	54.8	55.6
Current alcohol consumption (mean \pm SD) (g/day)*	23.9 \pm 28.5	27.0 \pm 28.3	25.3 \pm 28.5
BMI (kg/m ²) (mean \pm SD)	22.7 \pm 3.2	23.4 \pm 3.4	23.0 \pm 3.3
Smoking status			
Current smokers (%)	17.4	20.5	18.8
Former smokers (%)	24.0	22.4	23.3
Never smokers (%)	58.5	57.1	57.9

Table 1. Baseline characteristics of the study subjects. *Among current alcohol consumers.

revealed that rs2074356 achieved genome-wide significance ($P = 2.2 \times 10^{-16}$ for a meta-analysis). These findings indicate that the 12q24.12-13 locus is associated with coffee consumption among a Japanese population.

Coffee is one of the most widely consumed beverages worldwide¹. Recent national data from Japan have revealed that the average per capita consumption of coffee is about 11 cups/week². The role of coffee in human health has received much attention³. In prospective cohort studies and meta-analysis studies, coffee consumption has been inversely associated with risk of stroke, cardiovascular disease, and multiple chronic diseases, such as Parkinson disease, diabetes, and liver, urine, prostate and colorectal cancers⁴⁻¹⁰. For most populations, coffee is one of the most highly caffeinated beverages consumed. Given that there is considerable inter-individual variability in preference for caffeine, it has been suggested that habitual caffeine consumption is influenced by genetic factors, in addition to cultural, psychosocial, or environmental factors (smoking)¹¹. Twin studies among populations of European ancestry reported heritability estimates for caffeine use which ranged from 36% to 58%¹². Five genome-wide association studies (GWAS) have been carried out on coffee or caffeine consumption¹³⁻¹⁷. The early GWASs discovered associations between coffee or caffeine consumption and several genes, namely *CYP1A1-CYP1A2*¹³⁻¹⁵, *AHR*^{13,14}, *NRCAM* and *ULK3*¹⁵. A very large GWAS among nearly 130 thousand people confirmed the association with *CYP1A1-CYP1A2* and *AHR*, and also identified six novel loci, namely *ABCG2*, *POR*, *BDNF*, *SLC6A4*, *MLXIPL*, and *GCKR*¹⁶. Most recently, a GWAS revealed a significant association between *PDSS2* and habitual coffee consumption¹⁷. All these studies investigated subjects of European and/or African American ancestry, however, and no study has been conducted in an Asian population.

Here, we conducted a genome-wide association study to identify common genetic variations that affect coffee consumption in a Japanese population.

Results

We analyzed the effects of common variants on coffee consumption from two study populations in the J-MICC study, namely 6,312 individuals for the discovery stage and 4,949 individuals for the replication stage. We also analyzed the 11,261 individuals (total of the two study populations) for the replication of SNPs previously reported in western populations. Baseline characteristics of these two groups of participants are shown in Table 1, and baseline characteristics of participants according to site are shown in Supplementary Table 1. Mean age in the discovery and replication populations was 53.0 \pm 9.9 and 55.1 \pm 8.8 years old, and percentage of female participants was 55% and 53%, respectively. Coffee consumption was 1.6 \pm 1.5 cups/day for the discovery and 1.7 \pm 1.5 cups/day for the replication populations.

Genome-wide association study in a Japanese population. *Discovery stage.* We performed a genome-wide scan for habitual coffee consumption-associated genetic variants based on discovery samples ($N = 6,312$) with adjustment for age and sex. The quantile-quantile plot of the observed P values is shown in Fig. 1. The inflation factor of the genome-wide scan was 1.002 (95% confidence interval: 1.001–1.004), indicating that the population structure was well-adjusted. Figure 2 shows scatter plots of P values derived from genome-wide scan results for coffee consumption, which found that two independent loci (12q24.12–13 and 5q33.3) met suggestive significance ($P < 1 \times 10^{-6}$) (Table 2). Genome-wide analyses adjusted for age, sex and smoking status and adjusted for age, sex, smoking status and BMI did not find any other loci achieving suggestive significance (Supplementary Tables 2 and 3). The association between 12q24.12–13 locus and habitual coffee consumption was not attenuated by modifying adjustment variables (Supplementary Tables 2 and 3).

Regarding the 12q24.12–13 locus, the strongest significance was observed at rs2074356, which is located at an intron of the *HECTD4* gene (Fig. 3A). The rs2074356 A allele was significantly associated with high consumption of coffee ($P = 1.8 \times 10^{-11}$), and its effect size was estimated as 0.20 (standard error = 0.03) cups/day per allele. A conditional analysis showed that the association between 12q24 variants and habitual coffee consumption did not achieve suggestive significance when conditioned on the rs2074356 genotype (Fig. 3B and Supplementary

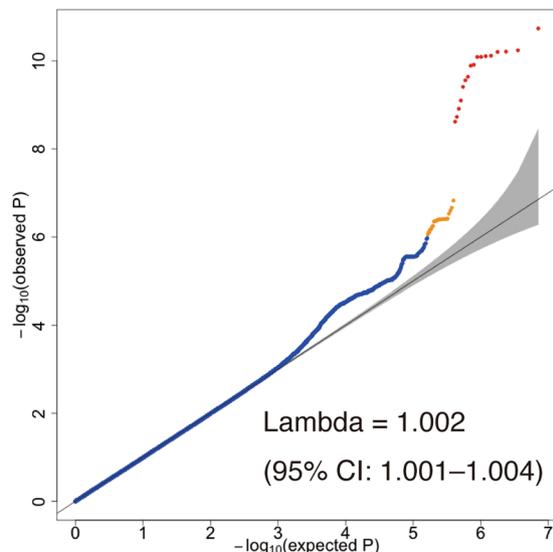


Figure 1. Quantile-quantile plot of genome-wide association tests using discovery samples ($N=6,312$). The x-axis indicates the expected $-\log_{10} P$ -values under the null hypothesis. The y-axis shows the observed $-\log_{10} P$ -values calculated by a mixed linear model association method. The black line represents $y=x$, which corresponds to the null hypothesis. The gray shaded area shows 95% confidence intervals of the null hypothesis. The inflation factor (lambda) is the median of the observed test statistics divided by the median of the expected test statistics. Variants with P -values indicating less than suggestive significance ($P < 1 \times 10^{-6}$) and genome-wide significance ($P < 5 \times 10^{-8}$) are shown in orange and red, respectively.

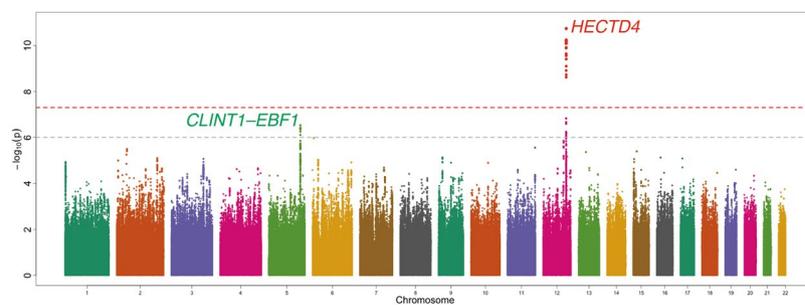


Figure 2. Genome-wide association signals from the discovery samples ($N=6,312$). The x-axis represents chromosomal positions and the y-axis represents $-\log_{10} P$ -values calculated by a mixed linear model association analysis. The grey and red dotted horizontal lines indicate the suggestive ($P=1 \times 10^{-6}$) and genome-wide ($P=5 \times 10^{-8}$) significance levels, respectively. Variants with P -values indicating less than genome-wide significance ($P < 5 \times 10^{-8}$) are shown in red.

Table 4). The frequency of the rs2074356 A allele was 25.2% in our discovery population. The rs2074356 A allele is East Asian-specific and monomorphic in Europeans, Africans, Americans, and South Asians according to the 1000 Genomes reference panel^{18,19}.

The lead variant at 5q33.3 was rs1957553, which is located at an intergenic region between *CLINT1* and *EBF1*. The rs1957553 A allele was associated with a 0.15 (SE = 0.03) cups/day per allele increase in habitual coffee consumption ($P=3.0 \times 10^{-7}$). The frequency of the rs1957553 A allele was 27.2% in our discovery population versus 61% in a European population of the 1000 Genomes reference panel^{18,19}.

Replication stage and meta-analysis. In the analysis of replication samples ($N=4,949$), rs2074356 was strongly associated with habitual coffee consumption ($P=2.2 \times 10^{-6}$), but no significant association was seen for rs1957553 ($P=0.53$). A meta-analysis of the discovery and replication populations revealed that rs2074356 achieved genome-wide significance ($P < 5 \times 10^{-8}$), but rs1957553 did not (Table 2). The phenotypic variance explained by rs2074346 was estimated at 0.59% from the meta-analysis.

In the meta-analysis, 24 variants in the 12q24.12–13 locus met genome-wide significance (Table 3). Of these 24 variants, 6 were located at intergenic regions; 15 were at intron regions; 1 was in the 3' UTR; 1 was a synonymous variant; and 1 was a missense variant. The 1 missense variant was rs671, which is located in the *ALDH2* gene and has been associated with alcohol drinking²⁰. We investigated the expression quantitative trait loci (eQTL)

SNP	Chr ^b	Gene(s)	Position ^c	EA ^d	NEA ^e	Rsq ^f	Population	AF ^g	Beta ^h	SE (Beta) ⁱ	Variance explained (%)	P
rs1957553	5	CLINT1-EBF1 (intergenic)	157,506,734	A	G	0.996	Discovery	0.272	0.1489	0.0290	0.42	3.0×10^{-7}
							Replication	0.276	-0.0216	0.0348	0.01	5.3×10^{-1}
							Meta-analysis	0.274	0.0789	0.0223	0.11	4.0×10^{-4}
rs2074356 ^a	12	HECTD4 (intron)	112,645,401	A	G	0.996	Discovery	0.252	0.2011	0.0299	0.73	1.8×10^{-11}
							Replication	0.224	0.1777	0.0376	0.44	2.2×10^{-6}
							Meta-analysis	0.240	0.1920	0.0234	0.59	2.2×10^{-16}

Table 2. SNPs associated with habitual coffee consumption. ^aDirectly genotyped; ^bChromosome; ^cChromosomal position (GRCh37/hg19); ^dEffect allele; ^eNon-effect allele; ^fImputation quality in terms of R-square calculated by the Minimac3 software version 1.0.11; ^gAllele frequency of effect allele; ^hEffect size; ⁱStandard error of effect size. Results listed in bold are the associations whose P-values are less than of genome-wide significance ($P < 5 \times 10^{-8}$).

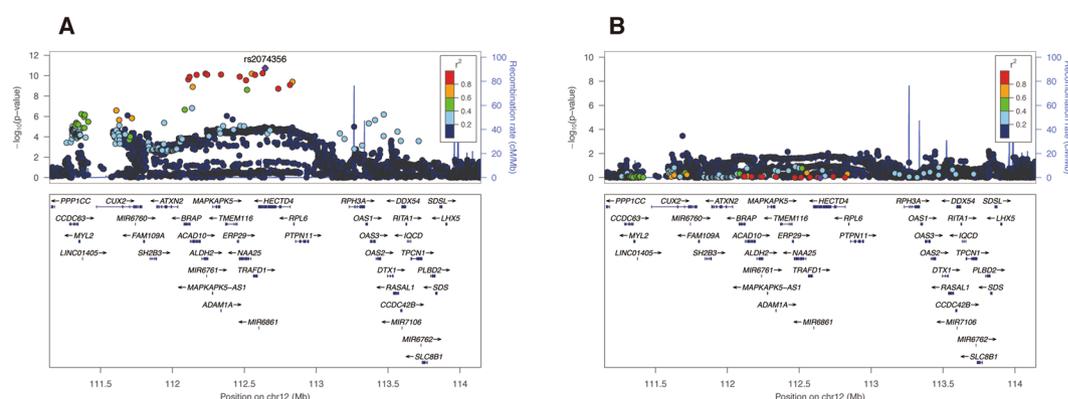


Figure 3. Association signals around the *HECTD4* gene using discovery samples ($N = 6,312$). The x-axis represents chromosomal positions near the *HECTD4* gene, and the y-axis represents $-\log_{10} P$ -values. The top signal in this locus (rs2074356) is shown in purple. Dot color for a variant represents the degree of linkage disequilibrium (R^2) estimates between each variant and rs2074356. (A) Signals from a genome-wide association scan adjusted for age and sex. (B) Signals from conditional analysis adjusted for age, sex and rs2074356 dosage.

relationship between the 24 significant variants and surrounding genes (Table 3). From the GTEx database²¹, no eQTL hit was found, possibly because the 12q24.12–13 variants are monomorphic in European populations. Accordingly, we looked up the Human Genetic Variation Database²², which is based on Japanese data, and found 5 genes (*TRAFD1*, *ALDH2*, *HECTD4*, *MAPKAPK5*, and *RPH3A*) whose expression levels were nominally significantly associated with the 12q24.12–13 variants ($P < 0.05$; Table 3).

Combined analysis of discovery and replication subjects. The above-mentioned analyses employed a discovery-replication scheme, which is useful for avoiding false positive associations caused by confounding factors, such as population stratification. We also investigated if additional loci associated with habitual coffee consumption might be suggested from our Japanese data using genome-wide association tests including both discovery and replication subjects ($N = 11,261$), with three sets of adjustment variables: (i) age and sex, (ii) age, sex, and smoking status, and (iii) age, sex, smoking status, and BMI. The results showed that only the 12q24.12–13 locus achieved genome-wide significance (Supplementary Table 5 and Supplementary Figure 1). An additional three loci had suggestive significance (Supplementary Table 5 and Supplementary Figure 1). Of these three loci, the *AGR3-AHR* locus had been associated with habitual caffeine or coffee consumption in previous GWASs^{13,14,16}. The other two loci, *CT49-DNAH5* and *MAB21L3-ATPIA1*, were not reported in previous GWASs, and were therefore considered to be novel candidate loci potentially associated with habitual coffee consumption.

Confounding factor adjustment. Rs 671, one of the significant 12q24 variants, is well known to be a functional polymorphism in the *ALDH2* gene that affects the activity of ALDH2 in East Asian populations²⁰. Reduced activity of ALDH2 is associated with increased concentrations of the toxin acetaldehyde and manifestation of the alcohol flush reaction, which protects individuals with the *ALDH2* 504Lys variant(s) from heavy drinking²³. We estimated the association between rs2074356 and coffee consumption adjusted for alcohol consumption in the 11,261 individuals (total of the two study populations). In the additive model, the rs2074356 A allele was significantly associated with high coffee consumption after adjustment for age, sex and alcohol consumption ($\beta = 0.176$, $P < 0.001$). We also estimated the association between rs2074356 and coffee consumption in the 11,261 individuals when stratified into alcohol drinkers and non-drinkers. In alcohol drinkers, the rs2074356 A allele was

SNP	Chr ^b	Position ^c	Gene(s)	EA ^d	NEA ^e	Rsq ^f	AF ^g	Beta ^h	SE (Beta) ⁱ	Variance explained (%)	P	eQTL (P < 0.05)
rs12227162	12	111,367,244	MYL2-CUX2 (intergenic)	T	C	0.974	0.201	0.1756	0.0252	0.44	3.2 × 10 ⁻¹²	no hit
rs149607519	12	111,389,437	MYL2-CUX2 (intergenic)	G	C	0.993	0.206	0.1682	0.0247	0.41	1.0 × 10 ⁻¹¹	no hit
rs148177611	12	111,390,454	MYL2-CUX2 (intergenic)	T	TAGAA	0.987	0.208	0.1672	0.0247	0.41	1.3 × 10 ⁻¹¹	no hit
rs3809297	12	111,609,727	CUX2 (intron)	T	G	0.872	0.247	0.1670	0.0248	0.46	1.7 × 10 ⁻¹¹	no hit
rs11065992	12	112,085,496	BRAP (intron)	C	T	0.804	0.461	0.1259	0.0225	0.35	2.3 × 10 ⁻⁸	no hit
rs3782886 ^a	12	112,110,489	BRAP (synonymous)	C	T	0.996	0.277	0.1808	0.0224	0.58	6.7 × 10 ⁻¹⁶	TRAFD1;ALDH2; HECTD4;MAPKAPK5
rs11066001	12	112,119,171	BRAP (intron)	C	T	0.984	0.273	0.1835	0.0226	0.59	4.4 × 10 ⁻¹⁶	no hit
rs60125993	12	112,136,208	ACAD10 (intron)	C	CT	0.906	0.502	0.1417	0.0209	0.44	1.2 × 10 ⁻¹¹	no hit
rs11066008	12	112,140,669	ACAD10 (intron)	G	A	0.828	0.342	0.1646	0.0232	0.54	1.5 × 10 ⁻¹²	no hit
rs11066015 ^a	12	112,168,009	ACAD10 (intron)	A	G	0.999	0.264	0.1829	0.0227	0.57	8.9 × 10 ⁻¹⁶	no hit
rs4646776	12	112,230,019	ALDH2 (intron)	C	G	0.997	0.264	0.1843	0.0227	0.58	4.4 × 10 ⁻¹⁶	no hit
rs671 ^a	12	112,241,766	ALDH2 (missense)SIFT: deleterious PolyPhen: possibly damaging	A	G	0.999	0.264	0.1840	0.0227	0.58	4.4 × 10 ⁻¹⁶	TRAFD1; MAPKAPK5; HECTD4
rs78069066	12	112,337,924	MAPKAPK5 (3' UTR)	A	G	0.964	0.268	0.1874	0.0230	0.61	4.4 × 10 ⁻¹⁶	TRAFD1; HECTD4; MAPKAPK5; ALDH2
rs11066132	12	112,468,206	NAA25 (intron)	T	C	0.929	0.260	0.1884	0.0236	0.60	1.3 × 10 ⁻¹⁵	no hit
rs116873087	12	112,511,913	NAA25 (intron)	C	G	0.929	0.263	0.1876	0.0235	0.60	1.3 × 10 ⁻¹⁵	no hit
rs11066150	12	112,518,803	NAA25 (intron)	A	G	0.802	0.454	0.1421	0.0225	0.44	2.7 × 10 ⁻¹⁰	no hit
rs147992802	12	112,552,274	NAA25-TRAFD1 (intergenic)	T	C	0.803	0.337	0.1762	0.0236	0.61	8.1 × 10 ⁻¹⁴	no hit
rs12231737	12	112,574,616	TRAFD1 (intron)	T	C	0.936	0.269	0.1894	0.0233	0.62	4.4 × 10 ⁻¹⁶	no hit
rs144504271	12	112,627,350	HECTD4 (intron)	A	G	0.955	0.265	0.1895	0.0231	0.62	2.2 × 10 ⁻¹⁶	no hit
rs2074356 ^a	12	112,645,401	HECTD4 (intron)	A	G	0.996	0.240	0.1920	0.0234	0.59	2.2 × 10 ⁻¹⁶	TRAFD1;MAPKAPK5
rs77768175	12	112,736,118	HECTD4 (intron)	G	A	0.818	0.244	0.1987	0.0256	0.64	8.9 × 10 ⁻¹⁵	no hit
rs11066280 ^a	12	112,817,783	HECTD4 (intron)	A	T	0.997	0.279	0.1742	0.0223	0.54	5.3 × 10 ⁻¹⁵	RPH3A; TRAFD1;MAPKAPK5
rs11537471	12	112,834,586	HECTD4-RPL6 (intergenic)	G	A	0.843	0.340	0.1702	0.0229	0.57	1.1 × 10 ⁻¹³	no hit
rs139144808	12	113,470,025	OAS2-DTX1 (intergenic)	TA	T	0.873	0.221	0.1433	0.0256	0.31	2.2 × 10 ⁻⁸	no hit

Table 3. Functional annotations for SNPs associated with habitual coffee consumption in 12q24 locus.

^aDirectly genotyped; ^bChromosome; ^cChromosomal position (GRCh37/hg19); ^dEffect allele; ^eNon-effect allele; ^fImputation quality in terms of R-square calculated by the Minimac3 software version 1.0.11; ^gAllele frequency of effect allele; ^hEffect size; ⁱStandard error of effect size.

significantly associated with high coffee consumption after adjustment for age, sex and alcohol consumption ($\beta = 0.357$, $p < 0.001$). In non-alcohol drinkers, the rs2074356 A allele was significantly associated with high coffee consumption after adjustment for age and sex ($\beta = 0.119$, $P < 0.001$). Previous studies suggested the association of rs2074356 in *HECTD4* with body mass index (BMI)²⁴. A recent GWAS proposed that the minor allele of the *HECTD4* variant rs2074356 is associated with a recused Thoracic-to-Hip ratio²⁵, which relates to BMI level. We estimated the association between rs2074356 and coffee consumption adjusted for BMI levels in the 11,261 individuals. The rs2074356 A allele was significantly associated with high coffee consumption after adjustment for age, sex and BMI levels ($P < 0.001$). A recent study found that the *ALDH2* 504Lys variant(s) is associated with smoking initiation²⁶. Smoking is a factor associated with caffeine consumption¹¹. We estimated the association between rs2074356 and coffee consumption adjusted for smoking status in the 11,261 individuals. The rs2074356 A allele was significantly associated with high coffee consumption after adjustment for age, sex and smoking status ($P < 0.001$). In a regression analysis adjusted for age, sex, alcohol, BMI levels and smoking status, the rs2074356 A allele was significantly associated with high consumption of coffee ($\beta = 0.147$, $P < 0.001$ for overall, $\beta = 0.32$, $P < 0.001$ for alcohol drinkers, $\beta = 0.092$, $P = 0.002$ for non-alcohol drinkers).

Replication of previously reported SNPs in the Japanese population. The 5 GWASs on coffee or caffeine consumption described to date¹³⁻¹⁷ have reported 18 SNPs (Supplementary Table 6). All 5 previous GWASs were conducted in individuals of European and/or African American ancestry. Variants on 7p21 (rs4410790 and rs6968554) and 15q24 (rs2470893 and rs2472297) were well-replicated, mainly in the European populations^{13,27,28}. In the J-MICC population of Japanese, in contrast, rs2470893 and rs2472297 were monoallelic. Variants on 6q21 showed very low minor allele frequencies (MAFs) (< 0.002). Table 4 shows the associations between the remaining 11 SNPs and habitual coffee consumption with adjustment for age and sex. Six variants (rs1260326, rs4410790, rs6968554, rs6968865, rs17685 and rs6265) were nominally significant ($P < 0.05$), while three variants on 7p21 (rs4410790, rs6968554 and rs6968865) were significant after multiple correction ($P < 0.05/11$). We estimate how much phenotypic variance in coffee consumption could be explained by the SNPs identified in Table 4. The explained variance ranged between 0.05% and 0.19%. The effect directions of all nominally significant variants were consistent with previous GWASs¹³⁻¹⁷ (Table 4).

SNP	Chr ^b	Position ^c	Gene(s)	EA ^d	NEA ^e	Rsq ^f	AF ^g	Beta ^h	SE (Beta) ⁱ	Variance explained (%)	P	Effect direction in European GWAS ^j
rs1260326^a	2	27,730,940	GCKR (missense) SIFT: tolerated PolyPhen: benign	C	T	0.996	0.442	0.0476	0.0207	0.05	0.02	+
rs1481012	4	89,039,082	ABCG2 (intron)	A	G	0.975	0.704	0.0339	0.0226	0.02	0.13	+
rs4410790^a	7	17,284,577	AGR3-AHR (intergenic)	C	T	0.996	0.375	0.0959	0.0207	0.19	3.7 × 10⁻⁶	+
rs6968554^a	7	17,287,106	AGR3-AHR (intergenic)	G	A	0.999	0.341	0.0908	0.0212	0.16	1.9 × 10⁻⁵	+
rs6968865^a	7	17,287,269	AGR3-AHR (intergenic)	T	A	1.000	0.340	0.0908	0.0212	0.16	1.9 × 10⁻⁵	+
rs7800944	7	73,035,857	MLXIPL (intron)	C	T	0.930	0.114	0.0430	0.0331	0.02	0.19	+
rs17685^a	7	75,616,105	POR (3' UTR)	A	G	0.999	0.363	0.0554	0.0209	0.06	0.008	+
rs382140 ^a	7	107,782,200	LAMB4-NRCAM (intergenic)	A	G	0.997	0.254	0.0302	0.0230	0.02	0.19	+
rs6265^a	11	27,679,916	BDNF (missense) SIFT: tolerated PolyPhen: probably damaging	C	T	1.000	0.592	0.0460	0.0206	0.05	0.03	+
rs6495122 ^a	15	75,125,645	CPLX3-ULK3 (intergenic)	A	C	1.000	0.808	0.0068	0.0257	0.00	0.79	—
rs9902453	17	28,349,095	EFCAB5 (intron)	A	G	0.983	0.338	0.0072	0.0218	0.00	0.74	—

Table 4. Replication analysis using the J-MICC samples for previously-reported SNPs. ^aDirectly genotyped; ^bChromosome; ^cChromosomal position (GRCh37/hg19); ^dEffect allele; ^eNon-effect allele; ^fImputation quality in terms of R-square calculated by the Minimac3 software version 1.0.11; ^gAllele frequency of effect allele; ^hEffect size; ⁱStandard error of effect size; ^jConsistent direction is denoted as ‘+’ and inconsistent direction is denoted as ‘-’. Results listed in bold are associations whose P-value is less than genome-wide significance ($P < 5 \times 10^{-8}$).

Discussion

In this study, we conducted the first GWAS on coffee intake in an Asian population. Participants were 6,312 individuals from a Japanese cohort study. Replication was attempted in another 4,949 individuals from the same cohort. A meta-analysis of the discovery and replication populations, we discovered that 24 novel SNPs on a 12q24 locus had genome-wide significance with habitual coffee consumption. The 24 SNPs associated with coffee intake were located at 13 genes, namely the *ALDH2*, *ACAD10*, *BRAP*, *ADAM1A*, *NAA25*, *TRAFD1*, *RPL6*, *MYL2*, *CUX2*, *OAS2*, *DTX1*, *MAPKAPK5* and *HECTD4* regions on the 12q24.12–13. Because these genes showed strong linkage disequilibrium, our results suggest that the 12q24.12–13 locus is responsible for variations in coffee consumption. We also confirmed an association between coffee intake and 6 SNPs previously reported in western populations.

Associations of the discovered genes with coffee consumption are intriguing but have not been studied well. One of these SNPs associated with coffee, rs 671, is a missense mutation and a functional Glu504Lys polymorphism in the *ALDH2* gene, namely a substitution of the Glu at codon position 504 with Lys, which affects the activity of ALDH2²⁰. The reduced activity of ALDH2, shown with the *ALDH2* 504Lys variant(s), contributes to increasing blood concentrations of the toxic acetaldehyde and exhibition of the alcohol flush reaction that protects individuals with the *ALDH2* 504Lys variant(s) from heavy drinking. One study found that coffee consumption was higher with the *ALDH2* 504Lys allele in Japanese men²⁹, which was consistent with our result. The *ALDH2* 504Lys variant(s) is associated with smoking initiation²⁶, and smoking is associated with caffeine consumption¹¹. Rs2074356 is located at an intron of the *HECTD4* gene. *HECTD4* may encode E3 ubiquitin protein ligase, which is a member of the ubiquitin ligase family. E3 ligase is involved in the final step in the ubiquitination cascade, catalyzing transfer of ubiquitin from an E2 enzyme to form a covalent bond with a substrate lysine³⁰. rs2074356 in *HECTD4* has been associated with drinking behavior in Han Chinese³¹. We confirmed that the association of rs2074356 in *HECTD4* with coffee was not attenuated by adjustment for alcohol consumption. We also confirmed that the association was not attenuated by adjustment for smoking status. All these discovered genes showed strong linkage disequilibrium. The results suggest that the association of the 12q24.12–13 locus with coffee consumption is not confounded by alcohol drinking or smoking status. Previous studies have reported associations of this 12q24 region with metabolic syndrome, thoracic-to-hip ratio²⁵, kidney function³², and BMI levels³³. Coffee consumption is also reported to be inversely associated with BMI levels³⁴. Our results indicate that the 12q24 region is independently associated with coffee consumption after adjustment for BMI level. Although our results do not allow us to conclude which is SNP is the most closely associated with coffee consumption, evidence from our GWAS suggests that the 12q24.12–13 locus is strongly associated with habitual coffee consumption. Because the SNPs of genes in this study exist only in East Asians, their association with coffee consumption in western populations must be rare. Identified SNPs are associated with the expression level of *ALDH2*, *HECTD4*, *TRAFD1*, *MAPKAPK5* and *RPH3A*. TRAFD1 (TRAF-Type Zinc Finger Domain Containing 1), encoded by *TRAFD1*, is a negative feedback regulator that controls excessive immune responses³⁵. MAPKAPK5 (Mitogen-Activated Protein Kinase-Activated Protein Kinase 5), encoded by *MAPKAPK5*, is a tumor suppressor and member of the serine/threonine kinase family³⁶. In response to cellular stress and proinflammatory cytokines, this kinase is activated through its phosphorylation by MAP kinases, including MAPK1/ERK, MAPK14/p38-alpha, and MAPK11/p38-beta³⁶. RPH3A encoded by *RPH3A* is thought to be an effector for RAB3A, which is a small GTP-binding protein that acts in neurotransmitter exocytosis³⁷. Genome-wide association tests using both discovery and

replication subjects identified three loci with suggestive significance, among which the *AGR3–AHR* locus was shown to be associated with habitual caffeine or coffee consumption in previous GWASs^{13,14,16}. The aryl hydrocarbon receptor (AHR), encoded by *AHR*, is a ligand-activated transcription factor that induces genes encoding CYP1A1 and CYP1A2³⁸, of which CYP1A2 is involved in the metabolism of widely used drugs and is a caffeine-metabolized enzyme³⁹.

In Japan, the most popular types of coffee are instant coffee, brewed coffee and canned coffee². Brewed coffee is made by brewing hot water with ground coffee beans. Brewing is most commonly done by drip or filter, and less commonly under pressure with an espresso machine. Several limitations of this study warrant mention. First, we did not evaluate details of coffee intake, such as cup size, use of caffeinated or decaffeinated coffee, or method of preparation (filtered or boiled). However, decaffeinated coffee and boiled coffee are very uncommon in Japan, and it was considered that assessment of the use and methods of coffee consumption and evaluation of their effects among Japanese would be uninformative. Second, because only a small number of participants (<5% of total participants) were cancer patients, they tended to underreport past coffee consumption as a result of decreased dietary intake. We minimized this limitation by asking these patients about their lifestyle when they were healthy or before their current symptoms developed. However, because more than 95% of the study participants were from a healthy general population, we consider that this study has external validity for the general Japanese population. Lastly, most of the functional effects of these coffee consumption-associated SNPs, including rs2074356, remain unclear. The functional relevance of the identified SNPs to coffee consumption remains to be determined. Therefore, our findings warrant further functional study to support the observed association between variants in the 12q24.12–13 locus and coffee consumption.

Coffee consumption is well known to be associated with a reduced risk of stroke, cardiovascular disease, Parkinson disease, diabetes, as well as liver, urine, prostate and colorectal cancers^{4–10}. However, the genetic factors associated with coffee have never been considered in the association of health benefits with coffee intake. Adjustment for the genetic factors found in this study should aid in establishing the association between coffee consumption and health benefits. Our study indicates the need for further research to evaluate the effect of genetic factors and coffee consumption on the relationship between coffee consumption and health outcomes.

In conclusion, we have discovered that the 12q24.12–13 locus is associated with coffee consumption among a Japanese population. This is the first report to identify a SNP for coffee consumption in an Asian population. Further studies are needed to investigate the biological mechanism that links the 12q24.12–13 locus and coffee consumption.

Methods

Study population. The GWAS was conducted in participants aged 35–69 years as a cross-sectional study within the Japan Multi-Institutional Collaborative Cohort (J-MICC) study. The 14,539 subjects of the J-MICC study were recruited from 12 different areas throughout Japan (Chiba, Okazaki, Shizuoka-Daiko, Takashima, Kyoto, Sakuragaoka, Aichi, Saga, Kagoshima, Tokushima, Fukuoka and Kyushu-KOPS) between 2004 and 2013. The 2,830 participants from two areas (Fukuoka and Kyushu-KOPS) were excluded from this study because the questionnaire on habitual coffee consumption in these areas was inconsistent with that used in the other 10 areas. Subjects who did not answer the questionnaire on habitual coffee consumption were also excluded. After quality control filtering (described below), a total of 11,261 participants were used in this study. For the discovery stage, we used the samples from the 6,312 participants from the 6 areas of Chiba, Okazaki, Shizuoka-Daiko, Takashima, Kyoto and Sakuragaoka. For the replication stage, we used the 4,949 participants from the 4 areas of Aichi, Saga, Kagoshima and Tokushima. The J-MICC study is a large cohort study launched in 2005 to confirm gene environment interactions in lifestyle-related disease. Details of the J-MICC Study have been reported elsewhere⁴⁰. Briefly, participants completed a questionnaire about lifestyle and medical information, and donated a blood sample at the time of the baseline survey. The J-MICC study participants included community citizens, first-visit patients to a cancer hospital and health check examinees. All participants in this study gave written informed consent, and the study protocol was approved by the Ethics Committees of Aichi Cancer Center, Nagoya University Graduate School of Medicine, and the other institutions participating in the J-MICC study. The present study was conducted according to the principles expressed in the World Medical Association Declaration of Helsinki.

Phenotype. The questionnaire for the J-MICC studies included questions on medical history, height, weight, family history (parents and siblings), smoking and drinking habits, dietary habits, sleeping habits, physical exercise and reproductive history. All exposures were collected using a scientifically validated self-administered questionnaire^{41,42}. The questionnaire was checked by trained staff to ensure completeness and consistency. Information on coffee was obtained in terms of frequency and intake from seven categories (never, <2 cups/week, 3–4 cups/week, 5–6 cups/week, 1–2 cup/day, 3–4 cups/day, ≥5 cups/day), for each of two types of coffee (drip, filter or instant) and (canned, plastic bottled, or carton). Canned coffee is a ready-to-drink canned coffee beverage which is very popular in Japan. The coffee categories were converted to cups/day by taking the median value of each category. Total coffee consumption was estimated as the sum amount of the two coffee types.

Genotyping and quality control filtering. Buffy coat fractions and DNA were prepared from blood samples and stored at –80 °C at the central J-MICC Study office. DNA was extracted from all buffy coat fractions using a BioRobot M48 Workstation (Qiagen Group, Tokyo, Japan) at the central study office. For the samples from two areas (Fukuoka and Kyushu-KOPS), DNA was extracted locally from samples of whole blood using an automatic nucleic acid isolation system (NA-3000, Kurabo, Co., Ltd. Osaka, Japan). The 14,539 study participants from the 12 areas of the J-MICC study, which includes the discovery and replication subjects,

were genotyped at RIKEN Center for Integrative Medicine Sciences using a HumanOmniExpressExome-8 v1.2 BeadChip array (Illumina Inc., San Diego, CA, USA). Twenty-six samples with inconsistent sex information between the questionnaire and an estimate from genotyping were excluded. The identity-by-descent method implemented in the PLINK 1.9 software^{43,44} identified 388 close relationship pairs (π -hat > 0.1875) and one sample of each pair was excluded. Principal component analysis (PCA)^{45,46} with a 1000 Genomes reference panel (phase 3)^{18,19} detected 34 subjects whose estimated ancestries were outside the Japanese population⁴⁷. These 34 samples were excluded. The remaining 14,091 samples all met a sample-wise genotype call rate criterion (≥ 0.99). SNPs with a genotype call rate < 0.98 and/or a Hardy-Weinberg equilibrium exact test P -value < 1×10^{-6} were removed, resulting in 873,254 autosomal variants. Of these, 298,644 variants with a low minor allele frequency (MAF) < 0.01 were excluded. This quality control filtering resulted in 14,091 individuals and 574,423 SNPs. Of the 14,091 samples, 6,312 were from the 6 areas for discovery analysis and 4,949 were from the 4 areas for replication analysis. The replication samples were subjected to genome-wide genotyping, followed by genome-wide imputation. However, only candidate novel loci identified during the discovery analysis were examined in the replication analysis to avoid the risk of identifying false-positive associations. A total of 11,261 samples from the 10 areas were also analyzed for replication of the previously reported SNPs in western populations. In addition to the discovery and replication design, we additionally conducted combined analysis of all the discovery and replication subjects ($N = 11,261$) to determine if our Japanese data might indicate additional loci associated with habitual coffee consumption.

Genotype imputation. Genotype imputation was performed using SHAPIT⁴⁸ and Minimac3⁴⁹ software based on the 1000 Genomes reference panel (phase 3)¹⁸. After genotype imputation, strict quality control filters were applied; namely, variants with an $R^2 < 0.8$ ⁵⁰ and a MAF < 0.01 were excluded, resulting in 7,094,228 variants.

Association tests between genetic variants and habitual coffee consumption. The association between genetic variants and quantitative habitual coffee consumption was tested using the mixed linear model association (MLMA) method⁵¹ with adjustment for age and sex. The mixed linear model uses adjustment covariates as fixed-effect variables and a genetic relationship matrix (GRM) as a variance-covariance matrix for random effects. To calculate the GRM, the genotyped SNPs were excluded using the quality-control criteria proposed in a previous study (genotype call rate ≥ 0.95 , Hardy-Weinberg exact test P -value ≥ 0.05 , and minor allele frequency ≥ 0.01)⁵², and the remaining 482,567 SNPs on autosomal chromosomes were used. Calculation of the GRM and genome-wide association tests were performed with the GCTA software⁵³ version 1.24.2. An advantage of the MLMA method over a linear regression method adjusted for principal components is its prevention of false positive associations due to population or relatedness structure⁵⁴. Because the Japanese population structure is not perfectly homogenous⁴⁷ and previous Japanese GWASs adjusted for principal components, they reported genomic inflation factor values which were slightly higher than expected (> 1.0)⁵⁵⁻⁵⁷. Accordingly, we chose the MLMA method to avoid the detection of false positive associations.

In the discovery stage, associations between all imputed variants and habitual coffee consumption was tested with adjustment for age and sex. This adjustment is consistent with 3 of 5 previous GWASs^{13,15,17}. For variants achieving suggestive significance ($P < 1 \times 10^{-6}$) in the discovery stage, the associations with habitual coffee consumption were examined in the replication stage. We then combined the resulting summary statistics from the discovery and replication stages by using a fixed-effect model and inverse-variance weighting method for meta-analysis⁵⁸. Variants achieving genome-wide significance ($P < 5 \times 10^{-8}$) in the meta-analysis were considered to be habitual coffee consumption-associated variants. In 2 of 5 previous GWASs^{14,16}, smoking status was used as an adjustment variable. Accordingly, we additionally adjusted for smoking status in the discovery, replication and meta-analysis as a sensitivity analysis. Furthermore, we conducted discovery, replication and meta-analysis with adjustment for age, sex, smoking status and BMI because coffee consumption was reported to be inversely associated with BMI level³⁴ and BMI can be a confounding factor of the genetic association with habitual coffee consumption.

For variants achieving genome-wide significance in the meta-analysis, we conducted conditional analysis based on the discovery population. In the conditional analysis, we tested the association between each variant and habitual coffee consumption by MLMA with adjustment for age, sex, and dosage of lead variant.

For replication analysis of previously reported SNPs, samples from the 10 areas were used for the association tests ($N = 11,261$).

Functional annotations. We examined genomic locations of variants identified in this study based on the Ensembl⁵⁹ and UCSC⁶⁰ genome browsers. For missense variants, we looked up the Ensembl genome browser for bioinformatics prediction results from SIFT⁶¹ and PolyPhen⁶². *Cis*-eQTL pairs of variants and genes were obtained from the GTEx²¹ and Human Genetic Variation databases²².

Confounding factor adjustment. Total alcohol consumption was estimated as the summed amount of pure alcohol consumption. The frequency of alcohol consumption was obtained in six categories (none, 1–3 times/month, 1–2 times/week, 3–4 times/week, 5–6 times/week, and everyday). Non-alcohol drinkers were defined as those who consumed alcohol none and 1–3 times/month. Alcohol drinkers were defined as those who consumed alcohol more than once/week. Smoking status was entered under the three categories of none, former, and current smoking. Multivariate linear regression analysis was used to test associations between SNP and coffee consumption with an additive model adjusted for age (continuous), sex, alcohol consumption (g/day) and BMI (continuous). Multivariate linear regression analysis was also used to test the association between SNP and coffee consumption with an additive model adjusted for age (continuous), sex, alcohol consumption (g/day) and BMI (continuous) according to alcohol drinking status (non-alcohol drinker and alcohol drinker). Values of $p < 0.05$

were considered statistically significant. The analyses were performed with Stata v. 14.1 (STATA Corporation, College Station, TX, USA).

Data Availability Statement. The datasets generated during and/or analysed during the current study are not publicly available due to ethical restriction, but are available from the co-author on reasonable request.

References

- Grigg, D. The worlds of tea and coffee: Patterns of consumption. *GeoJournal* **57**, 283–294 (2002).
- All J Coffee Association. *Coffee Market in Japan “Coffee Consumption by Type of Coffee”*, <http://coffee.ajca.or.jp/english> (2017).
- Cano-Marquina, A., Tarin, J. J. & Cano, A. The impact of coffee on health. *Maturitas* **75**, 7–21, <https://doi.org/10.1016/j.maturitas.2013.02.002> (2013).
- Kokubo, Y. *et al.* The impact of green tea and coffee consumption on the reduced risk of stroke incidence in Japanese population: the Japan public health center-based study cohort. *Stroke* **44**, 1369–1374 (2013).
- Sugiyama, K. *et al.* Coffee consumption and mortality due to all causes, cardiovascular disease, and cancer in Japanese women. *J Nutr* **140**, 1007–1013 (2010).
- Ding, M., Bhupathiraju, S. N., Chen, M., van Dam, R. M. & Hu, F. B. Caffeinated and decaffeinated coffee consumption and risk of type 2 diabetes: a systematic review and a dose-response meta-analysis. *Diabetes Care* **37**, 569–586, <https://doi.org/10.2337/dc13-1203> (2014).
- Bravi, F., Bosetti, C., Tavani, A., Gallus, S. & La Vecchia, C. Coffee reduces risk for hepatocellular carcinoma: an updated meta-analysis. *Clin Gastroenterol Hepatol* **11**, 1413–1421 e1411, <https://doi.org/10.1016/j.cgh.2013.04.039> (2013).
- Shimazu, T. *et al.* Coffee consumption and risk of endometrial cancer: a prospective study in Japan. *Int J Cancer* **123**, 2406–2410, <https://doi.org/10.1002/ijc.23760> (2008).
- Li, Q. *et al.* Coffee consumption and the risk of prostate cancer: the Ohsaki Cohort Study. *Br J Cancer* **108**, 2381–2389, <https://doi.org/10.1038/bjc.2013.238> (2013).
- Akter, S. *et al.* Coffee drinking and colorectal cancer risk: an evaluation based on a systematic review and meta-analysis among the Japanese population. *Jpn J Clin Oncol*. (2016).
- Brice, C. F. & Smith, A. P. Factors associated with caffeine consumption. *Int J Food Sci Nutr* **53**, 55–64 (2002).
- Yang, A., Palmer, A. A. & de Wit, H. Genetics of caffeine consumption and responses to caffeine. *Psychopharmacology (Berl)* **211**, 245–257, <https://doi.org/10.1007/s00213-010-1900-1> (2010).
- Sulem, P. *et al.* Sequence variants at CYP1A1-CYP1A2 and AHR associate with coffee consumption. *Hum Mol Genet.* **20**, 2071–2077 (2011).
- Cornelis, M. C. *et al.* Genome-wide meta-analysis identifies regions on 7p21 (AHR) and 15q24 (CYP1A2) as determinants of habitual caffeine consumption. *PLoS Genet.* **7**, e1002033 (2011).
- Amin, N. *et al.* Genome-wide association analysis of coffee drinking suggests association with CYP1A1/CYP1A2 and NRCAM. *Mol Psychiatry* **17**, 1116–1129, <https://doi.org/10.1038/mp.2011.101> (2012).
- Coffee and Caffeine Genetics Consortium. *et al.* Genome-wide meta-analysis identifies six novel loci associated with habitual coffee consumption. *Mol Psychiatry* **20**, 647–656, <https://doi.org/10.1038/mp.2014.107> (2015).
- Pirastu, N. *et al.* Non-additive genome-wide association scan reveals a new gene associated with habitual coffee consumption. *Sci Rep* **6**, 31590, <https://doi.org/10.1038/srep31590> (2016).
- Genomes Project, C. *et al.* An integrated map of genetic variation from 1,092 human genomes. *Nature* **491**, 56–65, <https://doi.org/10.1038/nature11632> (2012).
- Genomes Project, C. *et al.* A global reference for human genetic variation. *Nature* **526**, 68–74, <https://doi.org/10.1038/nature15393> (2015).
- Matsuo, K. *et al.* Gene-environment interaction between an aldehyde dehydrogenase-2 (ALDH2) polymorphism and alcohol consumption for the risk of esophageal cancer. *Carcinogenesis* **22**, 913–916 (2001).
- Consortium, G. T. Human genomics. The Genotype-Tissue Expression (GTEx) pilot analysis: multitissue gene regulation in humans. *Science* **348**, 648–660, <https://doi.org/10.1126/science.1262110> (2015).
- Higasa, K. *et al.* Human genetic variation database, a reference database of genetic variations in the Japanese population. *J Hum Genet* **61**, 547–553, <https://doi.org/10.1038/jhg.2016.12> (2016).
- Bosron, W. F. & L., T. K. Genetic polymorphism of human liver alcohol and aldehyde dehydrogenases, and their relationship to alcohol metabolism and alcoholism. *Hepatology* **6**, 502–510 (1986).
- Ninomiya-Baba, M. *et al.* Association of body mass index-related single nucleotide polymorphisms with psychiatric disease and memory performance in a Japanese population. *Acta Neuropsychiatr*, 1–10, <https://doi.org/10.1017/neu.2016.66> (2016).
- Cha, S., Park, A. Y. & Kang, C. A Genome-Wide Association Study Uncovers a Genetic Locus Associated with Thoracic-to-Hip Ratio in Koreans. *PLoS One* **10**, e0145220 (2015).
- Masaoka, H. *et al.* Combination of ALDH2 and ADH1B polymorphisms is associated with smoking initiation: A large-scale cross-sectional study in a Japanese population. *Drug Alcohol Depend.* **1**, 85–91 (2017).
- Josse, A. R., La, D. C., Campos, H. & El-Sohemy A. Associations between polymorphisms in the AHR and CYP1A1-CYP1A2 gene regions and habitual caffeine consumption. *Am J Clin Nutr* **96**, 665–671 (2012).
- McMahon, G., Taylor, A. E., Davey Smith, G. & Munafo, M. R. Phenotype refinement strengthens the association of AHR and CYP1A1 genotype with caffeine consumption. *PLoS One* **9**, e103448, <https://doi.org/10.1371/journal.pone.0103448> (2014).
- Yin, G. *et al.* ALDH2 polymorphism is associated with fasting blood glucose through alcohol consumption in Japanese men. *Nagoya J Med Sci.* **78**, 183–193 (2016).
- Berndsen, C. E. & Wolberger, C. New insights into ubiquitin E3 ligase mechanism. *Nat Struct Mol Biol.* **21**, 301–307 (2014).
- Yang, X. *et al.* Common variants at 12q24 are associated with drinking behavior in Han Chinese. *Am J Clin Nutr* **97**, 545–551 (2013).
- Okada, Y. *et al.* Meta-analysis identifies multiple loci associated with kidney function-related traits in east Asian populations. *Nat Genet* **44**, 904–909, <https://doi.org/10.1038/ng.2352> (2012).
- Wen, W. *et al.* Meta-analysis of genome-wide association studies in East Asian-ancestry populations identifies four new loci for body mass index. *Hum Mol Genet* **23**, 5492–5504, <https://doi.org/10.1093/hmg/ddu248> (2014).
- Grosso, G. *et al.* Association of daily coffee and tea consumption and metabolic syndrome: results from the Polish arm of the HAPIEE study. *Eur J Nutr* **54**, 1129–1137, <https://doi.org/10.1007/s00394-014-0789-6> (2015).
- Sanada, T. *et al.* FLN29 deficiency reveals its negative regulatory role in the Toll-like receptor (TLR) and retinoic acid-inducible gene I (RIG-I)-like helicase signaling pathway. *J Biol Chem* **283**, 33858–33864, <https://doi.org/10.1074/jbc.M806923200> (2008).
- Zhou, J. *et al.* MK5 is degraded in response to doxorubicin and negatively regulates doxorubicin-induced apoptosis in hepatocellular carcinoma cells. *Biochem Biophys Res Commun.* **427**, 581–586 (2012).
- Mizoguchi, A. *et al.* Localization of Rabphilin-3A on the synaptic vesicle. *Biochem Biophys Res Commun* **202**, 1235–1243, <https://doi.org/10.1006/bbrc.1994.2063> (1994).
- Nukaya, M., Moran, S. & Bradfield, C. A. The role of the dioxin-responsive element cluster between the Cyp1a1 and Cyp1a2 loci in aryl hydrocarbon receptor biology. *Proc Natl Acad Sci USA* **106**, 4923–4928 (2009).

39. Faber, M. S., Jetter, A. & Fuhr, U. Assessment of CYP1A2 activity in clinical practice: why, how, and when? *Basic Clin Pharmacol Toxicol.* **97**, 125–134 (2005).
40. Hamajima, N. & Group, J. M. S. The Japan Multi-Institutional Collaborative Cohort Study (J-MICC Study) to detect gene-environment interactions for cancer. *Asian Pac J Cancer Prev* **8**, 317–323 (2007).
41. Tokudome, S. *et al.* Development of a data-based short food frequency questionnaire for assessing nutrient intake by middle-aged Japanese. *Asian Pac J Cancer Prev* **5**, 40–43 (2004).
42. Tokudome, S. *et al.* Development of data-based semi-quantitative food frequency questionnaire for dietary studies in middle-aged Japanese. *Jpn J Clin Oncol* **28**, 679–687 (1998).
43. Purcell, S. *et al.* PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet* **81**, 559–575, <https://doi.org/10.1086/519795> (2007).
44. Chang, C. C. *et al.* Second-generation PLINK: rising to the challenge of larger and richer datasets. *Gigascience* **4**, 7, <https://doi.org/10.1186/s13742-015-0047-8> (2015).
45. Price, A. L. *et al.* Principal components analysis corrects for stratification in genome-wide association studies. *Nat Genet* **38**, 904–909, <https://doi.org/10.1038/ng1847> (2006).
46. Patterson, N., Price, A. L. & Reich, D. Population structure and eigenanalysis. *PLoS Genet* **2**, e190, <https://doi.org/10.1371/journal.pgen.0020190> (2006).
47. Yamaguchi-Kabata, Y. *et al.* Japanese population structure, based on SNP genotypes from 7003 individuals compared to other ethnic groups: effects on population-based association studies. *Am J Hum Genet* **83**, 445–456, <https://doi.org/10.1016/j.ajhg.2008.08.019> (2008).
48. Delaneau, O., Marchini, J. & Zagury, J. F. A linear complexity phasing method for thousands of genomes. *Nat Methods* **9**, 179–181, <https://doi.org/10.1038/nmeth.1785> (2011).
49. Das, S. *et al.* Next-generation genotype imputation service and methods. *Nat Genet* **48**, 1284–1287, <https://doi.org/10.1038/ng.3656> (2016).
50. Kiryluk, K. *et al.* GWAS for serum galactose-deficient IgA1 implicates critical genes of the O-glycosylation pathway. *PLoS Genet* **13**, e1006609, <https://doi.org/10.1371/journal.pgen.1006609> (2017).
51. Yang, J. *et al.* Common SNPs explain a large proportion of the heritability for human height. *Nat Genet* **42**, 565–569, <https://doi.org/10.1038/ng.608> (2010).
52. Lee, S. H., Wray, N. R., Goddard, M. E. & Visscher, P. M. Estimating missing heritability for disease from genome-wide association studies. *Am J Hum Genet* **88**, 294–305, <https://doi.org/10.1016/j.ajhg.2011.02.002> (2011).
53. Yang, J., Lee, S. H., Goddard, M. E. & Visscher, P. M. GCTA: a tool for genome-wide complex trait analysis. *Am J Hum Genet* **88**, 76–82, <https://doi.org/10.1016/j.ajhg.2010.11.011> (2011).
54. Yang, J., Zaitlen, N. A., Goddard, M. E., Visscher, P. M. & Price, A. L. Advantages and pitfalls in the application of mixed-model association methods. *Nat Genet* **46**, 100–106, <https://doi.org/10.1038/ng.2876> (2014).
55. Low, S. K. *et al.* Genome-wide association study of breast cancer in the Japanese population. *PLoS One* **8**, e76463, <https://doi.org/10.1371/journal.pone.0076463> (2013).
56. Urabe, Y. *et al.* A genome-wide association study of nephrolithiasis in the Japanese population identifies novel susceptible loci at 5q35.3, 7p14.3, and 13q14.1. *PLoS Genet.* <https://doi.org/10.1371/journal.pgen.1002541> (2012).
57. Okada, Y. *et al.* A genome-wide association study in 19 633 Japanese subjects identified LHX3-QSOX2 and IGF1 as adult height loci. *Hum Mol Genet.* **19**, 2303–2312 (2010).
58. Willer, C. J., Li, Y. & Abecasis, G. R. METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics* **26**, 2190–2191, <https://doi.org/10.1093/bioinformatics/btq340> (2010).
59. Aken, B. L. *et al.* Ensembl 2017. *Nucleic Acids Res* **45**, D635–D642, <https://doi.org/10.1093/nar/gkw1104> (2017).
60. Tyner, C. *et al.* The UCSC Genome Browser database: 2017 update. *Nucleic Acids Res* **45**, D626–D634, <https://doi.org/10.1093/nar/gkw1134> (2017).
61. Kumar, P., Henikoff, S. & Ng, P. C. Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. *Nat Protoc.* **4**, 1073–1081 (2009).
62. Adzhubei, I. A. *et al.* A method and server for predicting damaging missense mutations. *Nat Methods* **7**, 248–249, <https://doi.org/10.1038/nmeth0410-248> (2010).

Acknowledgements

We thank all the staff of the Laboratory for Genotyping Development, Center for the Integrative Medical Sciences, RIKEN, and the staff of the BioBank Japan project. This study was supported by Grants-in-Aid for Scientific Research for Priority Areas of Cancer (No. 17015018) and Innovative Areas (No. 221S0001) and by JSPS KAKENHI Grants (No. 16H06277 and 15H02524) from the Japanese Ministry of Education, Culture, Sports, Science and Technology. This work was also supported in part by a Grant-in-Aid from the Japan Society for the Promotion of Science (JSPS) KAKENHI Grant (Grant-in-Aid for Young Scientists) Number 15K19236 and 17K15840. This study was supported in part by funding for the BioBank Japan Project from the Japan Agency for Medical Research and development since April 2015, and the Ministry of Education, Culture, Sports, Science and Technology from April 2003 to March 2015.

Author Contributions

The authors' responsibilities were as follows – K.W. and H.T.: designed and supervised this research; and reviewed and edited the manuscript; H.S.N.: wrote the manuscript; and had primary responsibility for final content; A.S. and T.H.: analyzed data; and wrote the manuscript; M.N.: analyzed data; M.K. and Y.M.: performed GWAS genotyping; K.W., M.N., A.H., R.O., S.K. and T.S.: provided essential data set; M.H., Y.N., K.E., K.K., S.K.K., K.A., Y.N., R.I., S.S., A.H., H.M., Y.N., N.T., Y.N., N.K., E.O., N.F., H.I., H.S., I.O., M.W., K.M. and H.I.: conducted research (data collection); and all authors: read and approved the final manuscript.

Additional Information

Supplementary information accompanies this paper at <https://doi.org/10.1038/s41598-018-19914-w>.

Competing Interests: The authors declare that they have no competing interests.

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2018