Effect of epigenetic modulation on cancer sphere

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Abstract: Background: Cancer stem cell properties are highly relevant to the biology of treatment-resistant cancers. Epigenetic modification regulates gene expressions by chromatin remodeling during malignant transformation. The aim of this study was to elucidate the possible strategy for cancer stem cells focusing on epigenetic modification. Methods: We made cancer sphere from HepG2 cells, and we added Histone deacetylase (HDAC) inhibitor, valproic acid to cancer sphere. And we compared methylation status and the gene expression between normal HepG2 and cancer sphere groups, and between cancer sphere and sphere with HDAC inhibitor treatment groups. Results: Valproic acid (VPA) cancelled this spheroid formation. In comparison between normal HepG2 and cancer sphere, the number of methylation status changes more than 0.1 of beta level was 826 probes, and we could isolate some epithelial-mesenchymal transition (EMT) related genes. And VPA reduced the expressions of EMT related genes in sphere with RT-PCR. On the other hand, in comparison between cancer sphere and sphere with VPA treatment, we detected 29 probe of methylation status change, and VPA reduced the expressions of Bcl-6 in sphere. Conclusions: HDAC inhibitor affected the methylation status of cancer stem cells. Histone-acetylation might overcome treatment-resistant cancer through the regulation of cancer stem cell. J. Med. Invest. 67: 70-74, February, 2020

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INTRODUCTION

Hepatocellular carcinoma (HCC) is one of the most frequent cancers, and this is not only a leading cause of cancer-related deaths, but it is also significantly increasing globally (1,2). Despite short-term improvement afforded by surgical resection, local ablation therapy, transarterial chemoembolization, and liver transplantation, long-term outcome is still unsatisfactory in HCC patients (3-5).

Tumors contain the small subpopulations of cells called cancer stem cells (CSC) (6-9), and these cells are able to proliferate and self-renew extensively because of their ability to express anti-apoptotic and drug-resistant proteins, thus sustaining tumor growth (10).

Alterations in the epigenetic modulation of gene-expressions have been implicated in cancer development and progression. Histone acetylation is a post-translational modulation of the nucleosomal histones that affects chromatin structure and modulates gene-expressions. Histone deacetylases (HDACs) comprise an ancient family of enzymes that play crucial roles in numerous biological processes (11) and expressions of HDACs are upregulated in many tumor types (12,13). Therefore, HDAC inhibitors are promising as novel anticancer agents, and in this study we used Valproic acid (VPA), which has been shown to have antitumor effects in various cancer models (14-18).

And then, DNA methylation is observed in many human cancers, including HCC, in which global hypomethylation and specific promoter hypermethylation have been found as typical epigenetic changes involved in genomic instability and silencing of tumor suppressor genes, respectively (19,20). Regarding the relationship between epigenetic modification and cancer stem cell property, Nalls D et al. reported the possibility of epigenetic modification reduced cancer stem cell property and epigenetic modification may be promising reagents to boost the patient response to existing chemotherapies or as a standard cancer drug (21). However, there were numerous examples of aberrant DNA methylation, little is known about the global picture of hyper- or hypomethylated genes in CSC.

The aim of this study was to investigate the possible strategy for CSC through the epigenetic modification.

MATERIAL AND METHOD

Cell lines and culture conditions

HepG2 was provided by the RIKEN BRC through the National Bio-Resource Project of the MEXT, Japan. All cell lines were grown in RPMI 1640, supplemented with 10% fetal bovine serum (FBS), 70 μg/mL penicillin, 100 μg/mL streptomycin (complete medium), and maintained at 37°C in a humidified incubator with 5% CO2 in the air. The cells were maintained for no longer than 12 weeks after recovery from frozen stock.

Sphere formation

Cells were collected and washed to remove serum, then suspended in serum-free DMEM/F12 (Invitrogen) supplemented with 100 IU/ml penicillin, 100 μg/mL streptomycin, 20 ng/ml human recombinant epidermal growth factor (EGF) and 10 ng/ml human recombinant basic fibroblast growth factor (bFGF) (R&D systems, Minneapolis, MN, USA), 2% B27 supplement without vitamin A and 1% N2 supplement (Invitrogen, Carlsbad, CA, USA). The cells were subsequently cultured in ultra low attachment 6-well plates (Corning Inc., Corning, NY, USA) at a density of not more than 1 × 103 cells/well with or without 1.0 mM of VPA for 24 hours or 48 hours. The pictures of cells were taken by microscope (100x), (Inverted Microscope, Nikon TE2000-U, Japan)
Reagents

Valproic acid was purchased from Wako Pure Chemical Industries, Ltd. (Osaka, Japan), kept at 4°C, and diluted in PBS as necessary at the time of use.

Array-Based Analysis of DNA Methylation

Genomic DNA was extracted from HepG2 cell. Bisulfite conversion of 500 ng of genomic DNA was performed with the EZ DNA methylation kit (Zymo Research). DNA methylation level was assessed with Infinium® HumanMethylation450 BeadChips (Illumina Inc.). Quantitative measurements of DNA methylation were determined for 485,764 CpG dinucleotides that covered 99% of the RefSeq genes and were distributed across whole gene regions, including promoters, gene bodies, and 3′-UTRs. Detailed information on the contents of the array is available in the Infinium HumanMethylation450 User Guide, HumanMethylation450 manifest (www.illumina.com) and recent papers. DNA methylation data was analyzed using the methylation analysis module within the BeadStudio software (Illumina Inc.). DNA methylation statuses of the CpG sites were calculated as the ratio of the signal from a methylated probe relative to the sum of both methylated and unmethylated probes. These values, known as $\beta$-values, range from 0 (completely unmethylated) to 1 (fully methylated) (22,23). The genes in $\beta$-value difference $>0.1$ and $P < 0.05$ were detected.

And then, we used Ingenuity Pathway Analysis (IPA) 8.7 (http://www.ingenuity.com) to determine the functional pathways associated with the set of differentially expressed genes between genotypes. IPA utilizes the knowledge in the literature about biological interactions among genes and proteins.

RNA isolation and quantitative real-time RT-PCR

The extracted RNA was reversed, transcribed with High Capacity cDNA Reverse Transcription Kit (Applied Biosystems). Quantitative real-time RT-PCR was performed using the Applied Biosystems 7500 real-time PCR system, TaqMan Gene Expression Assays-on-demand, and the TaqMan Universal Master Mix (Applied Biosystems). The following assays (assay identification number) were used: CXCL5 (Hs01099660_g1), Bmi1 (Hs00180411_m1), Bcl-6 (Hs00153368_m1) and PIK3CA (Hs00907957_m1). TaqMan Human ACTB Endogenous Control (4326315E) was used as a control gene. The thermal cycler conditions were as follows: 2 minutes at 50°C, 10 minutes at 95°C, 40 cycles of 15 seconds at 95°C, and 1 minute at 60°C. Amplification data were analyzed with an Applied Biosystems Prism 7500 Sequence Detection System version 1.3.1 (Applied Biosystems).

Statistical analyses

Statistical comparisons of mean values were conducted using one-way ANOVA. All the results are presented as mean ± SD. All statistical analysis was performed using statistical software (JMP 8.0.1, SAS Campus Drive, Cary, NC). A $P$ value of less than 0.05 was considered to be statistically significant.

RESULTS

HepG2 well developed to cancer sphere, and VPA cancelled this spheroid formation with time dependent. 48 hr after VPA treatment, spheroid formation was completely cancelled (Fig. 1). In comparison between normal HepG2 and cancer sphere, the number of methylation status changes more than 0.1 of beta level was 826 probes. And hyper- and hypo-methylation changes were identified in 425 and 401 probe, respectively. Of these hyper- methylation changes, we could isolate some epithelial-mesenchymal transition (EMT) related genes like SDF-1, CXCL5 and IGFR1 (Fig. 2). In real-time RT-PCR, VPA reduced the CXCL5 expression compared to that of sphere with time ($P < 0.01$) (Fig. 3A). And regarding as other EMT related genes, Bmi1, we could get the similar data as CXCL5 (Fig. 3B). And we investigated the gene network with IPA analysis, IPA represented down-regulated (green) genes in the gene network of the “Tumor Development” in comparison between normal HepG2 and cancer sphere (Fig. 4).

On the other hand, in comparison between cancer sphere and sphere with VPA treatment, we detected 29 probe of methylation status change. And hyper- and hypo-methylation changes were identified in 20 and 9 probes, respectively. Especially in cell growth related gene, Bcl-6 changed to hyper- and PIK3CA changed to hypo-methylation status (Fig. 5).

In regard to the gene expressions, the down-regulation of the Bcl-6 expression in the sphere with VPA was confirmed by RT-PCR compared to the only sphere group with time ($P < 0.01$) (Fig. 6A). And then, the PIK3CA expression in the sphere with VPA tended to increase compared to that in only sphere group ($P = 0.08$) (Fig. 6B).

Figure 1: The morphological changes of HepG2 (A), sphere (B), sphere with VPA 24 hours (C) and sphere with VPA 48 hours (D)
Figure 2: The methylation status changes in comparison between normal HepG2 and cancer sphere.

Figure 3: The gene expressions of CXCL5 (A) and Bmi1 (B) with RT-PCR.

Figure 4: The gene network with the genes isolated in comparison between normal HepG2 and cancer sphere using IPA.

Figure 5: The methylation status changes in comparison between cancer sphere and sphere with VPA treatment.
DISCUSSION

In the present study, we assessed the epigenetic modulation in cancer stem cell, especially sphere formation with VPA on hepatoma cell line. To the best of our knowledge, this is the first report showing epigenetic modulation in cancer stem cell with methylation array.

In comparison between normal HepG2 and sphere, we could isolate some EMT related genes in hypo-methylation changes. The relationships between stemness characteristics and EMT phenomena have been reported in recent manuscripts. Researchers found that EMT acquires CSC properties, increases cancer cell tumorigenicity (24-26). In this study, VPA reduced the EMT related gene expressions and cancelled sphere formation, therefore these manuscripts are consistent with our findings.

In comparison between cancer sphere and sphere with VPA treatment, Bcl-6 changed to hyper- and PIK3CA changed to hypo-methylation status in cell growth related gene. Pellicano F et al. suggested leukemia cancer stem cell survival related to Bcl6 and PIK3CA, and these factors were inversely correlated (27). HDAC inhibitor, VPA seems to inhibit cancer stem cell survival through the epigenetic blocking in this pathway.

And then, we revealed HDAC inhibitor affected to methylation status changes. Zopf S et al. demonstrated that HDAC inhibitor reduced DNA methyltransferase (DNMT) activity and loss of DNMT activity was paralleled by a diminished methylation of the target gene in HepG2 cells (28). In other words, HDAC inhibitor can regulate the methylation status through the inhibition of DNMT.

VPA may have the effects to CSC in HepG2 cells in this study. However, some of the HDAC inhibitors are of limited therapeutic use due to the toxic side effects at high doses (29). VPA is widely used as a therapeutic drug for epilepsy and its toxic profile and pharmacokinetic properties are well established. Furthermore, in our study, the promising dose of VPA was 1.0 mM, while the peak plasma concentration in patients treated for epilepsy ranges between 0.5 and 1.2 mM (30). VPA, at a dose of 1.0 mM, may not cause any serious side effects in clinical setting.

In conclusion, VPA, one of the HDAC inhibitor, affected the methylation status of EMT related genes and cell growth related genes of cancer stem cells. Histone-acetylation might overcome treatment-resistant cancer through the regulation of cancer stem cell.

REFERENCES


We have no conflicts of interest or financial ties to disclose.

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Figure 6: The gene-expressions of Bcl-6 (A) and PIK3CA (B) with RT-PCR


