

RESEARCH ARTICLE

Elevated luminal inorganic phosphate suppresses intestinal Zn absorption in 5/6 nephrectomized rats

Yosuke Okumura,¹ Kotaro Abe,¹ Shoko Sakai,¹ Yuki Kamei,^{1,2} Yuki Mori,¹ Yuichiro Adachi,¹ Masaki Takikawa,¹ Ayano Kitamura,¹ Hirokazu Ohminami,¹ Kohta Ohnishi,¹ Masashi Masuda,¹ Taiho Kambe,³ Hironori Yamamoto,⁴ and Yutaka Taketani¹

¹Department of Clinical Nutrition and Food Management, Tokushima University Graduate School of Medical Nutrition, Tokushima, Japan; ²Department of Food and Nutrition, Tokushima University Graduate School of Medical Nutrition, Tokushima, Japan; ³Division of Integrated Life Science, Department of Applied Molecular Biology, Graduate School of Biostudies, Kyoto University, Kyoto, Japan; and ⁴Department of Health and Nutrition, Faculty of Human Life, Jin-ai University, Echizen, Japan

Abstract

Zinc (Zn) is an essential trace element in various biological processes. Chronic kidney disease (CKD) often leads to hypozincemia, resulting in further progression of CKD. In CKD, intestinal Zn absorption, the main regulator of systemic Zn metabolism, is often impaired; however, the mechanism underlying Zn malabsorption remains unclear. Here, we evaluated intestinal Zn absorption capacity in a rat model of CKD induced by 5/6 nephrectomy (5/6 Nx). Rats were given Zn and the incremental area under the plasma Zn concentration-time curve (iAUC) was measured as well as the expression of ZIP4, an intestinal Zn transporter. We found that 5/6 Nx rats showed lower iAUC than sham-operated rats, but expression of ZIP4 protein was upregulated. We therefore focused on other Zn absorption regulators to explore the mechanism by which Zn absorption was substantially decreased. Because some phosphate compounds inhibit Zn absorption by coprecipitation and hyperphosphatemia is a common symptom in advanced CKD, we measured inorganic phosphate (P_i) levels. P_i was elevated in not only serum but also the intestinal lumen of 5/6 Nx rats. Furthermore, intestinal intraluminal P_i administration decreased the iAUC in a dose-dependent manner in normal rats. In vitro, increased P_i concentration decreased Zn solubility under physiological conditions. Furthermore, dietary P_i restriction ameliorated hypozincemia in 5/6 Nx rats. We conclude that hyperphosphatemia or excess P_i intake is a factor in Zn malabsorption and hypozincemia in CKD. Appropriate management of hyperphosphatemia will be useful for prevention and treatment of hypozincemia in patients with CKD.

NEW & NOTEWORTHY We demonstrated that elevated intestinal luminal P_i concentration can suppress intestinal Zn absorption activity without decreasing the expression of the associated Zn transporter. Increased intestinal luminal P_i led to the formation of an insoluble complex with Zn while dietary P_i restriction or administration of a P_i binder ameliorated hypozincemia in chronic kidney disease model rats. Therefore, modulation of dietary P_i by P_i restriction or a P_i binder might be useful for the treatment of hypozincemia and hyperphosphatemia.

chronic kidney disease; dietary phosphate; hyperphosphatemia; zinc deficiency; zinc transporter

INTRODUCTION

Zinc (Zn) is an essential trace element that, as a cofactor of enzymes and transcription factors, maintains important biological processes such as cell division and differentiation (1). Hypozincemia is often a comorbidity of chronic kidney disease (CKD) (2). The prevalence of hypozincemia varies from 25% to 70% in patients on maintenance hemodialysis (3–5). Several factors affect serum Zn levels in patients with CKD. For instance, those are decreased intestinal absorption, decreased food intake, uremic toxicity, bioavailability, abnormal distribution to some tissues, increased fecal or urinary

loss, hemodialysis, aging, and multiple medications (6). Low dietary Zn intake is a significant risk factor for hypozincemia (7). On the other hand, serum Zn levels showed a negative relationship with the progression of the CKD stage (8, 9). In addition, hypozincemia in patients with CKD may increase the risk of end-stage kidney disease (10) and various complications such as hypertension, dyslipidemia, type II diabetes mellitus, taste disorders, cardiovascular disease including vascular calcification, and infectious disease (11–16). Therefore, prevention of hypozincemia is essential for delaying the progression of CKD and related complications and the decline in the patient's quality of life.

Zn supplementation is the standard treatment for hypozincemia and can generally increase serum Zn levels in patients with CKD (6). The dose of Zn supplementation is determined according to the severity of hypozincemia, ranging from 50 to 100 mg/day for adults and 1 to 3 mg/kg/day for children (6, 17). Current supplementation criteria do not consider the background and progression of the disease, which may lead to Zn under- or overdosing. In a randomized trial of patients on hemodialysis, for example, serum Zn levels were not sufficiently elevated even when standard doses of Zn were administered (18). Moreover, overdose of Zn often causes copper (Cu) and iron deficiency, leading to anemia and neurological disorders (19–22). At present, treatments that target the cause of hypozincemia are not well established in CKD.

The mechanism underlying hypozincemia in CKD has not been elucidated; however, increased urinary Zn excretion, decreased intestinal Zn absorption, and loss of Zn in dialysis have been proposed as potential causes (9, 23). Systemic Zn levels are mainly controlled by intestinal absorption. Cellular Zn flux is regulated by transporters, including members of SLC39A [Zrt-, Irt-like protein (ZIP)], and SLC30A [Zn transporter (ZnT)] families. ZIP4 (SLC39A4) is a responsible transporter of intestinal Zn absorption (24, 25). In addition to these transporter-mediated pathways, various intestinal luminal factors derived from the diet, including animal protein, sulfur-containing amino acids, calcium, and iron, have been reported to positively or negatively affect Zn absorption (26–29). Interestingly, phosphate compounds such as phytate (inositol hexaphosphate) and calcium phosphate can inhibit Zn absorption by coprecipitation with Zn (30, 31). In CKD, hyperphosphatemia, a common complication due to decreased urinary excretion of P_i , can increase secretion of P_i into the gastrointestinal tract (32, 33). Therefore, we hypothesized that elevated intestinal luminal P_i levels caused by hyperphosphatemia and/or dietary P_i intake may suppress intestinal Zn absorption in patients with CKD. In this study, we examined the mechanism underlying Zn malabsorption in a rodent model of CKD induced by 5/6 nephrectomy (5/6 Nx).

MATERIALS AND METHODS

Animals

All animals (male Wistar rats aged 5–8 wk) used in this study were purchased from Japan SLC, Inc. (Shizuoka, Japan). The animals were housed in a climate-controlled room ($22 \pm 2^\circ\text{C}$) under a 12:12-h light-dark cycle and maintained on standard chow (MF; P_i : 0.83%, Oriental Yeast, Tokyo, Japan) with free access to food and distilled water before induction of CKD or dietary intervention. All animals were euthanized under anesthesia between 9:00 AM and 12:30 PM for sample collection. All animal studies were approved by the animal experimentation committee of Tokushima University and were conducted in accordance with the guidelines for the management and handling of experimental animals.

5/6 Nx-Induced CKD Model

Five-week-old rats (Japan SLC) were randomly divided into a sham-operated group (hereafter sham rats) and a 5/6

Nx-induced CKD group (hereafter 5/6 Nx rats). Two-thirds of the left kidney mass of each 5/6 Nx rat was surgically removed under pentobarbital anesthesia (50 mg/kg ip). After 1 wk, the right kidney was removed under pentobarbital anesthesia. Sham operation was performed as described previously (34). 5/6 Nx rats were fed an MF diet (Oriental Yeast) or high- P_i MF diet (P_i : 1.2%, Oriental Yeast) for 9 wk and then fed an MF diet for 3 wk.

Measurement of Zn Uptake in Brush Border Membrane Vesicles

Brush border membrane vesicles (BBMVs) were prepared from the duodenum by using Ca^{2+} precipitation, as described previously (35, 36). Briefly, the mucosa of duodenum section was scraped, and homogenized in homogenate buffer (50 mM mannitol and 2 mM Tris-HCl buffer, pH 7.5). The homogenates were washed repeatedly by suspension and centrifugation in suspension buffer (300 mM mannitol and 10 mM Tris-HEPES, pH 7.5) to obtain purified BBMVs. All operations were performed at 4°C . A radioactivity assay was performed to analyze the uptake of ^{65}Zn (RIKEN, Saitama, Japan) into BBMVs by a rapid filtration technique (37). In brief, 10 μL of vesicle suspension was added to 90 μL of incubation solution (100 mM mannitol and 20 mM Tris-HEPES buffer, pH 7.0) containing ^{65}Zn (0.2 $\mu\text{Ci}/\text{mL}$), and the preparation was incubated at 20°C for 1 min. The reacted BBMV was filtrated on a nitrocellulose membrane and washed out free ^{65}Zn . The radioactivity [counts per minute (cpm)] of ^{65}Zn in the BBMV-bound membrane was measured by a gamma counter (Hitachi Aloka Medical, Tokyo, Japan). The Zn uptake activity (nmol/min) of BBMV was calculated from the radioactivity of BBMV divided by the specific activity of ^{65}Zn (cpm/nmol). The protein content of BBMV was determined by the Bradford method. Finally, Zn uptake activity was expressed by nmol/mg protein/min.

Plasma and Intestinal Intraluminal Fluid Biochemical Parameters

Blood samples were collected by inferior vena cava puncture into heparin-coated tubes (Mochida Pharmaceutical, Tokyo, Japan). After centrifugation for 15 min at 5,000 g, the supernatant was collected as plasma. The contents of the duodenum segment were extruded into 1.5-mL tubes. After centrifugation at 3,000 rpm for 5 min, the supernatant was collected as intestinal intraluminal fluid. The concentrations of Zn, P_i , calcium (Ca), and creatinine (Cre) in plasma and intestinal intraluminal fluid were determined by using a Metalloassay Kit (Metallogenics, Chiba, Japan), Phospha-C test (Wako, Osaka, Japan), Calcium-E test (Wako), and Labassay Creatinine (Wako), respectively. The concentration of blood urea nitrogen (BUN) was measured by Oriental Yeast.

Fecal Zn Excretion

Metabolic cages were used to collect fecal samples for 72 h before euthanasia. The feces were first dried at 110°C for 12 h and then micro pulverized, after which 100-mg samples were ashed at 250°C for 3 h, 350°C for 3 h, and 550°C for 24 h. These samples were heated at 100°C for 15 min with 10 mL of 1% HCl. Extracted Zn was measured using an inductively

coupled plasma optical emission spectrometer (ICP-OES; iCAP6300, Thermo Fisher Scientific, Waltham, MA). The fecal Zn excretion rate as the ratio of daily Zn excretion to daily Zn intake was then calculated because 5/6 Nx rats showed lower food intake (Supplemental Table S1).

Western Blot Analysis

Duodenal BBMVs were mixed in a sample buffer containing 5% 2-mercaptoethanol and subjected to SDS-PAGE. The separated proteins were transferred by electrophoresis to a polyvinylidene difluoride transfer membrane (Immobilon-P, Millipore), which was then treated with affinity-purified anti-ZIP4 (1:2,500) and mouse anti- β -actin monoclonal antibody (A5441, Sigma-Aldrich, St. Louis, MO, 1:5,000) as an internal control. Goat anti-mouse IgG (H + L)-HRP conjugate (1:4,000) was used as the secondary antibody, and signals were detected using Chemi-Lumi One Super (Nacalai Tesque, Kyoto, Japan) and luminescent image analyzer LAS-3000 (Fujifilm Life Science, Tokyo, Japan). Signal intensity was quantified with Multi Gauge V3.0 software (Fujifilm).

Real-Time PCR

Total RNA was isolated from harvested duodenum using RNAiso plus (Takara Bio, Shiga, Japan) and then digested by recombinant DNase I (Worthington Biochemical, Lakewood, NJ). First-strand cDNA was synthesized from 1.0 μ g of total RNA by using M-MLV reverse transcriptase (Nippon Gene, Tokyo, Japan), oligo (25) dT primer (Invitrogen), and dNTP mixture (Promega). After cDNA synthesis, real-time PCR was performed with the appropriate forward and reverse primers and Fast SYBR Green master mix (Applied Biosystems) using a real-time PCR system (StepOne Plus, Applied Biosystems). Real-time PCR primers (forward and reverse, respectively) were as follows: *Zip4* (*Slc39a4*), 5'-CAGCTACTGCAGAA-GATTGAGG-3' and 5'-CTTGGGAAGCAGGATCCATTAAG-3'; *Znt1* (*Slc30a1*), 5'-ACACGCTAGTGGCTAA-CACC-3' and 5'-AGACTGTCTGACTCCTGGATGA-3'; *Mt1*, 5'-TCTGTGCTT-ACACCGTTG-3' and 5'-AGCACTGTTCTGTCACCTCAG-3'; and *18 s rRNA*, 5'-GGGGAACGCGTGCATTTATC-3' and 5'-CTC-TCCGGAATCGAACCTG-3'. Using the comparative C_t method, each gene was quantified as mRNA level normalized to 18S rRNA.

Plasma Zn Concentration-Time Curve After Zn Administration

Total Zn absorption capacity was measured in sham, 5/6 Nx, and 8-wk-old normal rats. Sham rats and 5/6 Nx rats were fed an MF diet (Oriental Yeast) for 9 wk. Next, sham rats were fed an altered AIN-93G diet (Oriental Yeast) containing 0.6% P_i , and 5/6 Nx rats were fed an altered AIN-93G diet containing 1.2% P_i for 3 wk. A catheter (PE-50, Natsume Seisakusho, Tokyo, Japan) was inserted into the femoral vein of sham, 5/6 Nx, and normal rats under pentobarbital anesthesia. For sham and 5/6 Nx rats, 1 mL of 40 mg/mL $ZnSO_4$ solution was administered into the stomach by gavage, and plasma was collected at 0, 5, 10, 20, 30, and 40 min. For normal rats, 1 mL of 15 mM Zn solution was administered into the stomach by gavage without or with 15, 45, or 150 mM P_i and 3% lanthanum carbonate (La; No. 325767, Sigma-Aldrich), a potent binder of phosphate, and plasma

was collected at 0, 10, 20, 30, 40, and 60 min. P_i concentrations administered to the rats were selected to be equivalent to 3 or 10 times the intestinal luminal P_i levels in the sham rats. Zn levels were measured to obtain a plasma Zn concentration-time curve and the incremental area under the curve (iAUC) was determined.

Zn Solubility in the Presence of P_i

Zn solution (30 mM) was mixed with an equal volume of P_i solution (0–300 mM). The solution was passed through a 0.22- μ m filter (Wako). Zn in the filtrate was estimated as soluble Zn (31). Zn was analyzed by ICP-OES, and the solubility of Zn was calculated as the ratio of filtered Zn to Zn before filtration.

Effect of Dietary P_i Restriction and La on Hypozincemia

The effect of P_i restriction was measured in both 5/6 Nx- and adenine-induced CKD models. Sham and 5/6 Nx rats were fed an MF diet for 12 wk. Subsequently, 5/6 Nx rats were divided into a 5/6 Nx-control P_i (CP) group fed an altered AIN-93G diet (Oriental Yeast) containing 1.03% P_i (CP diet) and a 5/6 Nx-low P_i (LP) group fed an altered AIN-93G diet containing 0.2% P_i (LP diet) for 2 wk. Sham rats were fed a CP diet for 2 wk. Eight-week-old normal rats were randomly divided into a control group fed a CP diet for 40 days and an adenine group fed a CP diet containing 0.2–0.6% adenine (adenine-CP diet, A8626, Sigma-Aldrich) for 33 days. Subsequently, the adenine group was further divided into an adenine-CP group fed an adenine-CP diet and an adenine-LP group fed an LP diet containing 0.2–0.6% adenine for 7 days. In addition, 8-wk-old rats were randomly divided into a control group fed an MF diet and an adenine group fed an MF diet containing 0.4% adenine for 4 wk. The adenine group was then fed an MF diet containing 0.4% adenine with or without 6% La for 2 wk. Zn levels were measured in plasma samples as described above in *Plasma and Intestinal Intraluminal Fluid Biochemical Parameters*.

Statistical Analysis

Data are presented as means \pm SE. GraphPad PRISM software (version 5, GraphPad Software, San Diego, CA) was used for all statistical analyses. For comparison between two groups, a two-tailed unpaired *t* test was used. For multiple comparisons, one-way ANOVA followed by Tukey's tests was used. *P* values of <0.05 were considered statistically significant.

RESULTS

Imbalance in Zn Flux in the Intestine of 5/6 Nx Rats

In 5/6 Nx rats, plasma P_i , BUN, and Cre levels were significantly elevated, and Zn and Ca levels were significantly decreased compared with sham rats (Table 1), similar to the trend in biochemical parameters observed in patients with CKD. To assess intestinal Zn flux, we verified Zn excretion into feces. The fecal Zn content was increased, but there was no difference in daily fecal Zn excretion between sham and 5/6 Nx rats (Table 1). However, the fecal Zn excretion rate was significantly higher in 5/6 Nx rats than in sham rats (Table 1). Therefore, we hypothesized that Zn absorption is substantially decreased in CKD.

Table 1. Comparison of plasma biochemical parameters and fecal Zn excretion between Sham and 5/6 Nx rats

Biochemical Parameter	Sham	5/6 Nx
Plasma		
Zn, µg/dL	102 ± 15.5	58.9 ± 3.67*
Pi, mg/dL	6.52 ± 0.45	10.2 ± 0.87*
Ca, mg/dL	8.99 ± 0.26	6.51 ± 0.18*
BUN, mg/dL	17.3 ± 0.65	93.1 ± 10.8*
Cre, mg/dL	0.79 ± 0.09	1.83 ± 0.11*
Fecal Zn ¹		
Content, mg/g	0.24 ± 0.01	0.27 ± 0.01*
Excretion, mg/day	0.56 ± 0.03	0.53 ± 0.03
Excretion rate, %	63.4 ± 1.85	74.6 ± 1.41*

¹Amount of Zn, daily Zn excretion, and daily Zn excretion rate (Zn excretion/Zn intake) in feces were measured. Data are expressed as means ± SE (n = 5–7). Statistical analysis was performed using a two-tailed unpaired t test. *P < 0.05. BUN, blood urea nitrogen; Pi, inorganic phosphate. n indicates the number of samples for each experiment.

Enhanced Zn Transport by Duodenal BBMVs in 5/6 Nx Rats

ZIP4, a ZIP family Zn transporter, mainly localizes to the intestinal brush border membrane (BBM) and plays a central role in intestinal Zn absorption (38). Zn is most actively absorbed from the jejunum in humans (39) and the duodenum in rats (40); therefore, we evaluated the uptake of Zn by the duodenum BBM using a ⁶⁵Zn radioisotope. In 5/6 Nx rats, the Zn uptake activity of BBMVs was significantly increased compared with sham rats, contrary to our hypothesis (Fig. 1A). We also measured the expression of ZIP4, which is usually expressed as a 75-kDa protein. Under Zn deficiency, however, ZIP4 is intracellularly processed into a 37-kDa protein that accumulates on the BBM and upregulates Zn absorption from the diet (38, 41–43). Consistent with the activation of Zn uptake, ZIP4 protein levels were significantly elevated in BBMVs from 5/6 Nx rats (Fig. 1, B and C).

Zn is transported from the cytoplasm into the blood by ZnT1, which localizes to the basolateral membrane (BLM) of enterocytes (44). Intracellular Zn is stored in a form bound to metallothionein (MT), a Zn-binding protein (45). We found that mRNA levels of *Znt1* and *Mt1* in the duodenum were significantly decreased in 5/6 Nx rats compared with sham rats (Fig. 2, A and B). Both *Znt1* and *Mt1* gene expression have been reported to depend on dietary and intracellular Zn levels (46–48). These results suggest that enterocytes may not take up Zn.

Suppression of Elevated Plasma Zn Levels After Zn Administration in 5/6 Nx Rats

To evaluate the total Zn absorption capacity in 5/6 Nx rats, including transporters and intestinal luminal factors, we calculated the iAUC after Zn administration. Plasma Zn levels were significantly lower in 5/6 Nx rats than in sham rats at all timepoints after Zn administration (Fig. 3A). The iAUC also decreased in 5/6 Nx rats (Fig. 3B), suggesting that total Zn absorption capacity is reduced in 5/6 Nx-induced CKD.

Decreased iAUC After Zn Administration by P_i-Induced Reduction of Zn Solubility

To determine the cause of decreased total Zn absorption in 5/6 Nx rats, we focused on the intestinal intraluminal environment. We observed that intestinal intraluminal P_i

levels in sham and 5/6 Nx rats were 15.0 ± 1.89 and 22.1 ± 2.14 mM, respectively (Fig. 4A). To examine the effect of intestinal intraluminal P_i on Zn absorption, we evaluated the iAUC after Zn administration with or without P_i in normal rats, which showed that the iAUC decreased in a P_i concentration-dependent manner (Fig. 4, B and C). Similar to the known properties of Zn and P_i (49), the solubility of Zn was decreased by the concentrations of P_i used in this study (Fig. 4D). Furthermore, La, a potent binder of phosphate, significantly ameliorated the P_i-induced reduction in the iAUC (Fig. 5, A and B). Collectively, these results suggest that elevated intestinal intraluminal P_i in CKD inhibits Zn absorption by reducing Zn solubility.

Amelioration of Hypozincemia in CKD Rats by Dietary P_i Restriction and P_i Binder

Lastly, we examined whether dietary interventions other than Zn supplementation might ameliorate hypozincemia. We found that 5/6 Nx rats fed an LP diet had significantly higher plasma Zn levels compared with 5/6 Nx rats fed a CP

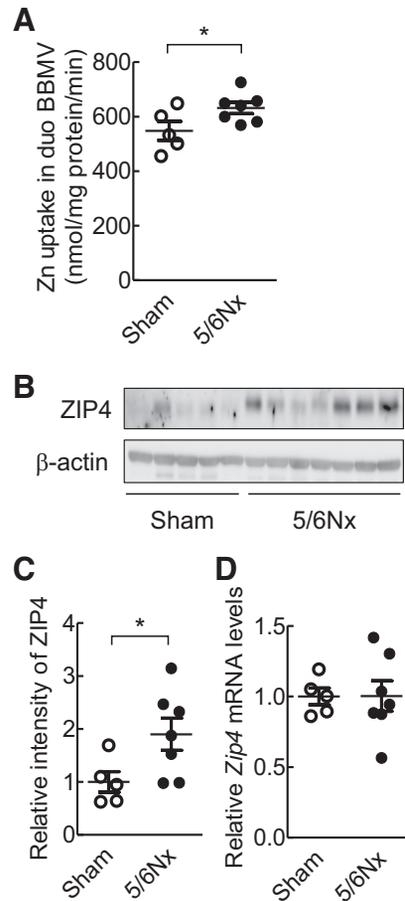


Figure 1. Elevated Zn transport activity in the duodenal BBMVs of 5/6 Nx rats. A: measurement of Zn transport activity by uptake of ⁶⁵Zn in duodenal BBMVs from sham rats (open circles) and 5/6 Nx rats (closed circles). B: Western blot of ZIP4 in duodenal BBMVs. Each lane was loaded with 20 µg of BBMVs. C: Western blot band intensities. ZIP4 protein levels were normalized to β-actin levels. 5/6 Nx intensities were normalized to Sham values. D: measurement of *Zip4* (*Slc39a4*) mRNA levels in the duodenum by real-time RT-PCR. Data are expressed as means ± SE (n = 5–7). Statistical analysis was performed using a two-tailed unpaired t test. *P < 0.05. BBMVs, brush border membrane vesicles. n indicates the number of samples for each experiment.

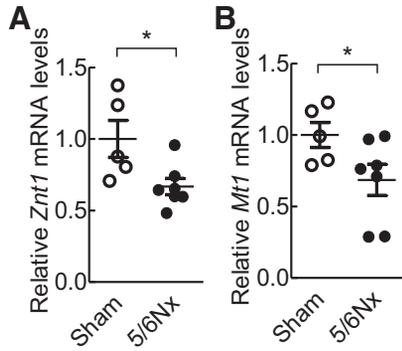


Figure 2. mRNA expression of *Znt1* and *Mt1* in the duodenum. Relative mRNA levels of *Znt1* (*Slc30a1*; A) and *Mt1* (B) in the duodenum of sham rats (open circles) and 5/6 Nx rats (closed circles) were measured by real-time RT-PCR. Data are expressed as means \pm SE ($n = 5-7$). Statistical analysis was performed using a two-tailed unpaired *t* test. * $P < 0.05$. *n* indicates the number of samples for each experiment.

diet (Fig. 6A). In addition, the LP diet significantly ameliorated the decrease in plasma Zn levels observed in adenine-induced CKD rats (Fig. 6B). Furthermore, administration of La improved hypozincemia in adenine-induced CKD rats (Fig. 6C). These results suggest that appropriate Zn supplementation, dietary P_i restriction, and P_i binder may also be effective for improving hypozincemia in CKD.

DISCUSSION

This study has demonstrated that elevated intestinal luminal P_i concentration suppresses intestinal Zn absorption in 5/6 Nx rats, a model of CKD. One of the mechanisms of Zn

malabsorption was a reduction of Zn solubility by excess intestinal luminal P_i . We showed that both solubility and absorption of Zn were decreased even under the condition of a 1:1 molar ratio of Zn to P_i . We found that the basal intestinal intraluminal P_i levels were ~ 1.5 times higher in 5/6 Nx rats than in sham rats. These results suggest that P_i -mediated suppression of Zn absorption can occur in a physiological environment and may be one of the causes of hypozincemia in CKD.

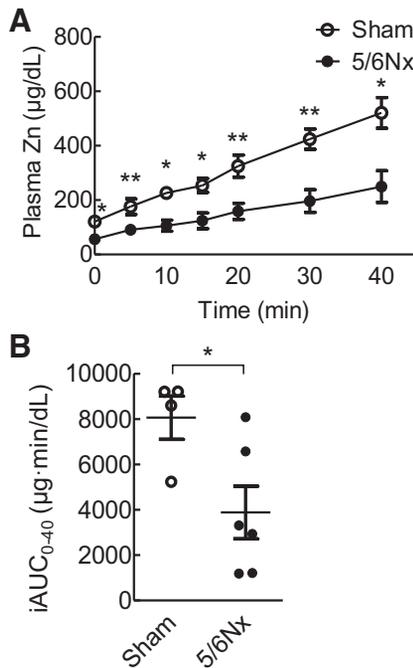


Figure 3. Low plasma Zn levels after Zn administration in 5/6 Nx rats. A: plasma Zn levels in sham rats (open circles) and 5/6 Nx rats (closed circles) at 0, 5, 10, 15, 20, 30, and 40 min after Zn administration. B: incremental area under the plasma Zn concentration-time curve (iAUC) in Sham and 5/6 Nx rats. Data are expressed as means \pm SE ($n = 4-6$). Statistical analysis was performed using a two-tailed unpaired *t* test. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$. *n* indicates the number of samples for each experiment.

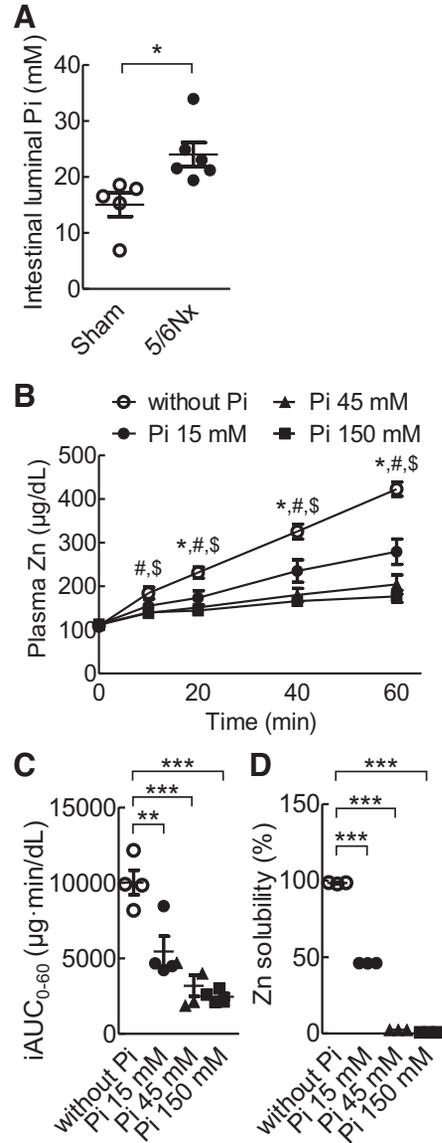


Figure 4. Inhibition of Zn absorption and decrease in Zn solubility by excess P_i . A: intestinal intraluminal P_i levels in the duodenum of Sham (open circle) and 5/6 Nx (closed circle) rats. Data are expressed as means \pm SE ($n = 5$ or 6). Statistical analysis was performed using a two-tailed unpaired *t* test. * $P < 0.05$. B: plasma Zn levels at 0, 10, 20, 30, 40, and 60 min after Zn administration. Eight-week-old rats were administered Zn solution without P_i (open circles) or containing P_i at 15 mM (closed circles), 45 mM (closed triangles), or 150 mM (closed squares). Statistical analysis was performed using one-way ANOVA followed by Tukey's tests. * $P < 0.05$ vs. P_i 15 mM; # $P < 0.05$ vs. P_i 45 mM; \$ $P < 0.05$ vs. P_i 150 mM. C: iAUC. D: solubility of Zn in the mixed solution (15 mM Zn and 15, 45, or 150 mM P_i) evaluated as the ratio of Zn pre- and post-filtration. Data are expressed as means \pm SE ($n = 3$ or 4). Statistical analysis was performed using one-way ANOVA followed by Tukey's tests. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$. iAUC, incremental area under the plasma Zn concentration-time curve. *n* indicates the number of samples for each experiment.

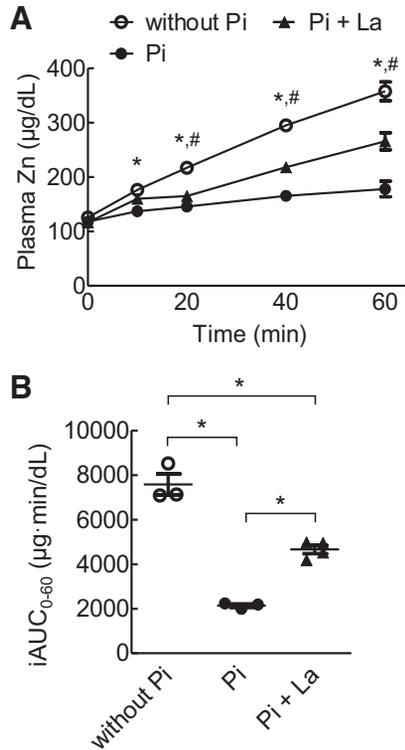


Figure 5. Lanthanum carbonate improved the P_i -induced reduction of Zn absorption. **A:** plasma Zn levels in 8-wk-old rats at 0, 10, 20, 30, 40, and 60 min after Zn administration with or without 45 mM P_i and 3% La. * $P < 0.05$ vs. P_i ; # $P < 0.05$ vs. $P_i + La$. **B:** incremental area under the plasma Zn concentration-time curve (iAUC) in the rats. Data are expressed as means \pm SE ($n = 3$ or 4). * $P < 0.05$. All statistical analysis was performed using one-way ANOVA followed by Tukey's tests. P_i , inorganic phosphate. n indicates the number of samples for each experiment.

In general, ZIP4 protein is upregulated to increase intestinal Zn absorption in situations of Zn deficiency or hypozincemia. We found that total intestinal Zn absorption was still suppressed in 5/6 Nx rats, although hypozincemia occurred and ZIP4 expression in the duodenum was elevated. These changes suggest that P_i causes intestinal Zn malabsorption by the reduction of Zn solubility and consequently induces the compensatory response to Zn deficiency in 5/6 Nx rats. Although the Zn compounds used in Zn supplement therapy, such as gluconate, citrate, and acetate, are readily dissociated and absorbed in the gastrointestinal tract, P_i can form an insoluble complex with Zn (49). We found that administration of La ameliorated the P_i -mediated reduction of Zn absorption. Collectively, these results suggest that suppression of Zn absorption in CKD may be, in part, induced by the formation of a Zn- P_i insoluble complex in the intestinal lumen.

Thus, our study has indicated that elevated P_i levels in the intestine negatively affect intestinal Zn absorption independently of the Zn transport activity of the duodenal BBM. However, there are several limitations in our study. First, we did not directly examine the amount of Zn- P_i complex formed in the intestinal lumen. Because the ease of formation of the Zn- P_i complex under CKD conditions is unknown, further studies will be needed to assess its contribution to the progression of hypozincemia in patients with CKD. Second, we did not evaluate the Zn transport activity of the

intestinal BLM in 5/6 Nx rats. We found that the mRNA levels of *Znt1* were decreased in 5/6 Nx rats, but its protein levels remain unclear. A recent study reported that ZnT1 protein levels are downregulated by hepcidin (50), a peptide hormone secreted from the liver, and increased in CKD. Furthermore, it has been reported that albumin on the basolateral side enhances Zn absorption in a coculture of intestinal cell line Caco-2 and the mucin-producing goblet cell line HT-29-MTX (51). Thus, Zn transport by the BLM may be involved in the downregulation of total Zn absorption in CKD. In our study, total Zn absorption in 5/6 Nx rats was decreased to about half that in sham rats, and we consider that an increase in P_i levels in the intestine is not the only mechanism that downregulates intestinal Zn absorption in

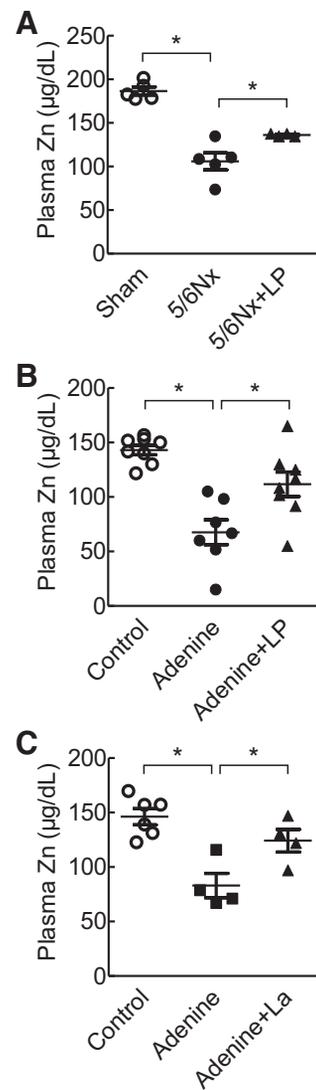


Figure 6. Amelioration of hypozincemia in CKD model rats by dietary P_i restriction and La. **A:** plasma Zn levels in 5/6 Nx rats fed a CP diet or LP diet for 2 wk. **B:** plasma Zn levels in adenine-induced CKD rats fed a CP diet or LP diet containing adenine for 7 days. **C:** plasma Zn levels in adenine-induced CKD rats fed an MF diet containing 0.4% adenine with or without La for 2 wk. Data are expressed as means \pm SE ($n = 4-8$). Statistical analysis was performed using one-way ANOVA followed by Tukey's tests. * $P < 0.05$. CKD, chronic kidney disease. n indicates the number of samples for each experiment.

CKD. Third, we evaluated the changes in plasma Zn levels after Zn administration as a surrogate index of total Zn absorption capacity. Plasma Zn levels reflect not only intestinal absorption but also kidney excretion and distribution to organs. In this study, the urinary excretion and distribution to organs of Zn would be less than the Zn absorption in the intestine, because plasma Zn concentration was linearly elevated at least for 40 min after Zn administration. Therefore, we consider that the changes in plasma Zn levels would reflect total Zn absorption capacity. Lastly, our study was based on rodent models of 5/6 Nx- and adenine-induced CKD, and the corresponding CKD stage of these models is not entirely clear. Hyperphosphatemia occurs more frequently in CKD stage 5 and dialysis (52). In a cohort study of patients with CKD, the fractional excretion of Zn into urine increased strongly at stage 3, suggesting that hypozincemia in CKD is not compensated by reduced renal Zn excretion (9). Thus, there may be different mechanisms underlying hypozincemia at each stage of CKD, and the actual contribution of P_i management to blood Zn levels in patients with CKD remains unclear. Further clinical investigation will be needed.

Supplementation with Zn is a useful treatment for hypozincemia, but several clinical issues remain to be resolved. Because Zn and Cu compete for absorption in the intestine, continuous administration of high-dose Zn often causes Cu deficiency and its related symptoms (21). Serum Zn and Cu levels have been found to be inversely correlated in patients with CKD (22). Our study showed that dietary P_i restriction and administration of La prevent a decrease in plasma Zn levels in CKD model rats, suggesting these approaches to hyperphosphatemia may be useful treatments that avoid the problem of current Zn supplementation therapy in hypozincemia. In this study, we did not evaluate the effect of P_i management on the expression of Zn transporters. Additional studies will be needed to elucidate the detailed mechanisms of improved hypozincemia by P_i management including Zn transporters.

In conclusion, an elevation in intestinal luminal P_i concentration due to hyperphosphatemia or excess P_i intake may suppress Zn absorption and cause hypozincemia in CKD. Dietary P_i restriction and administration of La can ameliorate hypozincemia in CKD model rats. Appropriate management of hyperphosphatemia might be useful for the prevention and treatment of hypozincemia in patients with CKD.

Perspectives and Significance

Zn plays a pivotal role in various biological processes. CKD often leads to hypozincemia, which is a risk factor for the progression and complications of CKD. The current treatment for hypozincemia is Zn supplementation, which often causes Cu deficiency. We demonstrated that suppression of Zn absorption in CKD may be, in part, induced by the formation of a Zn- P_i insoluble complex in the intestinal lumen. In addition, dietary P_i restriction can ameliorate hypozincemia in a rat model of CKD. Appropriate management of hyperphosphatemia may be useful for prevention and treatment of hypozincemia in patients with CKD, although further clinical investigation will be needed to clarify completely the

relationship between dietary P_i and Zn absorption capacity or hypozincemia in this disorder.

DATA AVAILABILITY

Data will be made available upon reasonable request.

SUPPLEMENTAL DATA

Supplemental Table S1: <https://doi.org/10.6084/m9.figshare.24234079>.

ACKNOWLEDGMENTS

We thank Aika Yoshizawa, Miho Samori, Ayano Komatsubara, Yutaro Nobe, and Mako Endo (Department of Clinical Nutrition and Food Management, Tokushima University Graduate School of Medical Nutrition, Tokushima, Japan) for technical assistance. We also thank Support Center for Advanced Medical Sciences, Tokushima University Graduate School of Biomedical Sciences, and Advance Radiation Research, Education, and Management Center, Tokushima University.

GRANTS

This work was supported by JSPS KAKENHI Grants 19H04053, 23H03329, 23KJ1663, and JST, the establishment of university fellowships toward the creation of science technology innovation, Grant JPMJFS2130. This work was also supported by Research Clusters of Tokushima University: Research Cluster for Precision Nutrition.

DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the authors.

AUTHOR CONTRIBUTIONS

Y.O. and Y.T. conceived and designed research; Y.O., K.A., S.S., Y.M., M.T., and M.M. performed experiments; Y.O., K.A., and S.S. analyzed data; Y.O., Y.K., Y.M., Y.A., M.T., A.K., H.O., K.O., M.M., T.K., and H.Y. interpreted results of experiments; Y.O. and K.A. prepared figures; Y.O. and Y.T. drafted manuscript; Y.O. and Y.T. edited and revised manuscript; Y.O. and Y.T. approved final version of manuscript.

REFERENCES

- Chasapis CT, Spiliopoulou CA, Loutsidou AC, Stefanidou ME. Zinc and human health: an update. *Arch Toxicol* 86: 521–534, 2012. doi:10.1007/s00204-011-0775-1.
- Elgenidy A, Amin MA, Awad AK, Husain-Syed F, Aly MG. Serum zinc levels in chronic kidney disease patients, hemodialysis patients, and healthy controls: systematic review and meta-analysis. *J Ren Nutr* 33: 103–115, 2023. doi:10.1053/j.jrn.2022.04.004.
- Shimizu S, Tei R, Okamura M, Takao N, Nakamura Y, Oguma H, Maruyama T, Takashima H, Abe M. Prevalence of zinc deficiency in Japanese patients on peritoneal dialysis: comparative study in patients on hemodialysis. *Nutrients* 12: 764, 2020. doi:10.3390/nu12030764.
- Bozalioğlu S, Ozkan Y, Turan M, Simşek B. Prevalence of zinc deficiency and immune response in short-term hemodialysis. *J Trace Elem Med Biol Organ Biol* 18: 243–249, 2005. doi:10.1016/j.jtemb.2005.01.003.
- Erten Y, Kayataş M, Sezer S, Özdemir FN, Özyiğit PF, Turan M, Haberal A, Güz G, Kaya S, Bilgin N. Zinc deficiency: prevalence and causes in hemodialysis patients and effect on cellular immune

- response. *Transplant Proc* 30: 850–851, 1998. doi:10.1016/S0041-1345(98)00075-X.
6. Nakatani S, Mori K, Shoji T, Emoto M. Association of zinc deficiency with development of CVD events in patients with CKD. *Nutrients* 13: 1680, 2021. doi:10.3390/nu13051680.
 7. Joo YS, Kim HW, Lee S, Nam KH, Yun H-R, Jhee JH, Han SH, Yoo T-H, Kang S-W, Park JT. Dietary zinc intake and incident chronic kidney disease. *Clin Nutr* 40: 1039–1045, 2021. doi:10.1016/j.clnu.2020.07.005.
 8. Shih CT, Shiu YL, Chen CA, Lin HY, Huang YL, Lin CC. Changes in levels of copper, iron, zinc, and selenium in patients at different stages of chronic kidney disease. *Genomic Med Biomark Health Sci* 4: 128–130, 2012. doi:10.1016/j.gmbhs.2013.03.001.
 9. Damianaki K, Lourenco JM, Braconnier P, Ghobril J-P, Devuyst O, Burnier M, Lenglet S, Augsburger M, Thomas A, Pruijm M. Renal handling of zinc in chronic kidney disease patients and the role of circulating zinc levels in renal function decline. *Nephrol Dial Transplant* 35: 1163–1170, 2020. doi:10.1093/ndt/gfz065.
 10. Tokuyama A, Kanda E, Itano S, Kondo M, Wada Y, Kadoya H, Kidokoro K, Nagasu H, Sasaki T, Kashiwara N. Effect of zinc deficiency on chronic kidney disease progression and effect modification by hypoalbuminemia. *PLoS One* 16: e0251554, 2021. doi:10.1371/journal.pone.0251554.
 11. Fukasawa H, Furuya R, Kaneko M, Nakagami D, Ishino Y, Kitamoto S, Omata K, Yasuda H. Clinical significance of trace element zinc in patients with chronic kidney disease. *J Clin Med* 12: 1667, 2023. doi:10.3390/jcm12041667.
 12. Ume AC, Wenegeime T-Y, Adams DN, Adesina SE, Williams CR. Zinc deficiency: a potential hidden driver of the detrimental cycle of chronic kidney disease and hypertension. *Kidney360* 4: 398–404, 2023. doi:10.34067/KID.0007812021.
 13. Tsutsumi R, Ohashi K, Tsutsumi YM, Horikawa YT, Minakuchi J, Minami S, Harada N, Sakae H, Sakai T, Nakaya Y. Albumin-normalized serum zinc: a clinically useful parameter for detecting taste impairment in patients undergoing dialysis. *Nutr Res* 34: 11–16, 2014. doi:10.1016/j.nutres.2013.10.009.
 14. Choi S, Liu X, Pan Z. Zinc deficiency and cellular oxidative stress: prognostic implications in cardiovascular diseases review-article. *Acta Pharmacol Sin* 39: 1120–1132, 2018. doi:10.1038/aps.2018.25.
 15. Ari E, Kaya Y, Demir H, Ascioglu E, Keskin S. The correlation of serum trace elements and heavy metals with carotid artery atherosclerosis in maintenance hemodialysis patients. *Biol Trace Elem Res* 144: 351–359, 2011. doi:10.1007/s12011-011-9103-0.
 16. Saka Y, Naruse T, Matsumoto J, Takeda Y, Onogi C, Yokoi J, Kato A, Tawada N, Noda Y, Niwa S, Mimura T, Watanabe Y. Low serum zinc concentration is associated with infection particularly in patients with stage 5 chronic kidney disease medicated with proton pump inhibitors. *J Ren Nutr* 31: 579–585, 2021. doi:10.1053/j.jrn.2020.11.006.
 17. Kodama H, Tanaka M, Naito Y, Katayama K, Moriyama M. Japan's practical guidelines for zinc deficiency with a particular focus on taste disorders, inflammatory bowel disease, and liver cirrhosis. *Int J Mol Sci* 21: 2941, 2020. doi:10.3390/ijms21082941.
 18. Tonelli M, Wiebe N, Thompson S, Kinniburgh D, Klarenbach SW, Walsh M, Bello AK, Faruque L, Field C, Manns BJ, Hemmelgarn BR; Alberta Kidney Disease Network. Trace element supplementation in hemodialysis patients: a randomized controlled trial. *BMC Nephrol* 16: 52, 2015. doi:10.1186/s12882-015-0042-4.
 19. Prasad AS, Rabbani P, Brewer GJ, Schoomaker EB. Hypocupremia induced by zinc therapy in adults. *JAMA* 240: 2166–2168, 1978. doi:10.1001/jama.1978.03290200044019.
 20. Yadrick MK, Kenney MA, Winterfeldt EA. Iron, copper, and zinc status: response to supplementation with zinc or zinc and iron in adult females. *Am J Clin Nutr* 49: 145–150, 1989. doi:10.1093/ajcn/49.1.145.
 21. Duncan A, Yacoubian C, Watson N, Morrison I. The risk of copper deficiency in patients prescribed zinc supplements. *J Clin Pathol* 68: 723–725, 2015. doi:10.1136/jclinpath-2014-202837.
 22. Nishime K, Kondo M, Saito K, Miyawaki H, Nakagawa T. Zinc burden evokes copper deficiency in the hypoalbuminemic hemodialysis patients. *Nutrients* 12: 577, 2020. doi:10.3390/nu12020577.
 23. Mahajan SK, Bowersox EM, Rye DL, Abu-Hamdan DK, Prasad AS, McDonald FD, Biersack KL. Factors underlying abnormal zinc metabolism in uremia. *Kidney Int Suppl* 27: S269–S273, 1989.
 24. Kambe T, Tsuji T, Hashimoto A, Isumura N. The physiological, biochemical, and molecular roles of zinc transporters in zinc homeostasis and metabolism. *Physiol Rev* 95: 749–784, 2015. doi:10.1152/physrev.00035.2014.
 25. Maares M, Haase H. A guide to human zinc absorption: general overview and recent advances of in vitro intestinal models. *Nutrients* 12: 762, 2020. doi:10.3390/nu12030762.
 26. Heth DA, Becker WM, Hoekstra WG. Effect of calcium, phosphorus and zinc on zinc-65 absorption and turnover in rats fed semipurified diets. *J Nutr* 88: 331–337, 1966. doi:10.1093/jn/88.3.331.
 27. Miquel E, Alegria A, Barberá R, Farré R. Casein phosphopeptides released by simulated gastrointestinal digestion of infant formulas and their potential role in mineral binding. *Int Dairy J* 16: 992–1000, 2006. doi:10.1016/j.idairyj.2005.10.010.
 28. Krebs NF. Overview of zinc absorption and excretion in the human gastrointestinal tract. *J Nutr* 130: 1374S–1377S, 2000. doi:10.1093/jn/130.5.1374S.
 29. Yang Y, Zhu S, Guo W, Feng Y, Guo T, Wu H. Formation of calcium phosphate nanoparticles mediated by animal protein hydrolysates enhances calcium absorption by murine small intestine: ex vivo. *Food Funct* 10: 6666–6674, 2019. doi:10.1039/c9fo01273g.
 30. Lonnerdal B, Sandberg AS, Sandstrom B, Kunz C. Inhibitory effects of phytic acid and other inositol phosphates on zinc and calcium absorption in suckling rats. *J Nutr* 119: 211–214, 1989. doi:10.1093/jn/119.2.211.
 31. Feng Y, Zhang J, Miao Y, Guo W, Feng G, Yang Y, Guo T, Wu H, Zeng M. Prevention of zinc precipitation with calcium phosphate by casein hydrolysate improves zinc absorption in mouse small intestine ex vivo via a nanoparticle-mediated mechanism. *J Agric Food Chem* 68: 652–659, 2020. doi:10.1021/acs.jafc.9b07097.
 32. Taketani Y, Koiba F, Yokoyama K. Management of phosphorus load in CKD patients. *Clin Exp Nephrol* 21: 27–36, 2017. doi:10.1007/s10157-016-1360-y.
 33. Savica V, Calò L, Santoro D, Monardo P, Granata A, Bellinghieri G. Salivary phosphate secretion in chronic kidney disease. *J Ren Nutr* 18: 87–90, 2008. doi:10.1053/j.jrn.2007.10.018.
 34. Ghosh SS, Massey HD, Krieg R, Fazalbhoy ZA, Ghosh S, Sica DA, Fakhry I, Gehr TWB. Curcumin ameliorates renal failure in 5/6 nephrectomized rats: role of inflammation. *Am J Physiol Renal Physiol* 296: F1146–F1157, 2009. doi:10.1152/ajprenal.90732.2008.
 35. Taketani Y, Segawa H, Chikamori M, Morita K, Tanaka K, Kido S, Yamamoto H, Iemori Y, Tatsumi S, Tsugawa N, Okano T, Kobayashi T, Miyamoto K, Takeda E. Regulation of type II renal Na⁺-dependent inorganic phosphate transporters by 1,25-dihydroxyvitamin D₃. Identification of a vitamin D-responsive element in the human NAPI-3 gene. *J Biol Chem* 273: 14575–14581, 1998. doi:10.1074/jbc.273.23.14575.
 36. Masuda M, Yamamoto H, Takei Y, Nakahashi O, Adachi Y, Ohnishi K, Ohminami H, Yamanaka-Okumura H, Sakae H, Miyazaki M, Takeda E, Taketani Y. All-trans retinoic acid reduces the transcriptional regulation of intestinal sodium-dependent phosphate co-transporter gene (Npt2b). *Biochem J* 477: 817–831, 2020. doi:10.1042/BCJ20190716.
 37. Gunshin H, Noguchi T, Naito H. Effect of calcium on the zinc uptake by brush border membrane vesicles isolated from the rat small intestine. *Agric Biol Chem* 55: 2813–2816, 1991. doi:10.1271/BBB1961.55.2813.
 38. Weaver BP, Dufner-Beattie J, Kambe T, Andrews GK. Novel zinc-responsive post-transcriptional mechanisms reciprocally regulate expression of the mouse Slc39a4 and Slc39a5 zinc transporters (Zip4 and Zip5). *Biol Chem* 388: 1301–1312, 2007. doi:10.1515/BC.2007.149.
 39. Lee HH, Prasad AS, Brewer GJ, Owyang C. Zinc absorption in human small intestine. *Am J Physiol Gastrointest Liver Physiol* 256: G87–G91, 1989. doi:10.1152/ajpgi.1989.256.1.G87.
 40. Davies NT. Studies on the absorption of zinc by rat intestine. *Br J Nutr* 43: 189–203, 1980. doi:10.1079/bjn19800078.
 41. Kambe T, Andrews GK. Novel proteolytic processing of the ectodomain of the zinc transporter ZIP4 (SLC39A4) during zinc deficiency is inhibited by acrodermatitis enteropathica mutations. *Mol Cell Biol* 29: 129–139, 2009. doi:10.1128/MCB.00963-08.
 42. Hashimoto A, Nakagawa M, Tsujimura N, Miyazaki S, Kizu K, Goto T, Komatsu Y, Matsunaga A, Shirakawa H, Narita H, Kambe T, Komai M. Properties of Zip4 accumulation during zinc deficiency

- and its usefulness to evaluate zinc status: a study of the effects of zinc deficiency during lactation. *Am J Physiol Regul Integr Comp Physiol* 310: R459–R468, 2016. doi:10.1152/ajpregu.00439.2015.
43. **Hoadley JE, Leinart AS, Cousins RJ.** Kinetic analysis of zinc uptake and serosal transfer by vascularly perfused rat intestine. *Am J Physiol Gastrointest Liver Physiol* 252: G825–G831, 1987. doi:10.1152/ajpgi.1987.252.6.g825.
 44. **Yu YY, Kirschke CP, Huang L.** Immunohistochemical analysis of ZnT1, 4, 5, 6, and 7 in the mouse gastrointestinal tract. *J Histochem Cytochem* 55: 223–234, 2007. doi:10.1369/jhc.6A7032.2006.
 45. **Krežel A, Maret W.** The functions of metamorphic metallothioneins in zinc and copper metabolism. *Int J Mol Sci* 18: 1237, 2017. doi:10.3390/ijms18061237.
 46. **McMahon RJ, Cousins RJ.** Regulation of the zinc transporter ZnT-1 by dietary zinc. *Proc Natl Acad Sci USA* 95: 4841–4846, 1998. doi:10.1073/pnas.95.9.4841.
 47. **Laity JH, Andrews GK.** Understanding the mechanisms of zinc-sensing by metal-response element binding transcription factor-1 (MTF-1). *Arch Biochem Biophys* 463: 201–210, 2007. doi:10.1016/j.abb.2007.03.019.
 48. **Hennigar SR, Kelley AM, McClung JP.** Metallothionein and zinc transporter expression in circulating human blood cells as biomarkers of zinc status: a systematic review. *Adv Nutr* 7: 735–746, 2016. doi:10.3945/an.116.012518.
 49. **Speight JG, Lange NA, Dean JA.** *Lange's Handbook of Chemistry* (16th ed). New York: McGraw-Hill, 2005.
 50. **Hennigar SR, McClung JP.** Hepcidin attenuates zinc efflux in Caco-2 cells. *J Nutr* 146: 2167–2173, 2016. doi:10.3945/jn.116.237081.
 51. **Maares M, Duman A, Keil C, Schwerdtle T, Haase H.** The impact of apical and basolateral albumin on intestinal zinc resorption in the Caco-2/HT-29-MTX co-culture model. *Metallomics* 10: 979–991, 2018. doi:10.1039/c8mt00064f.
 52. **Oka T, Hamano T, Sakaguchi Y, Yamaguchi S, Kubota K, Senda M, Yonemoto S, Shimada K, Matsumoto A, Hashimoto N, Mori D, Monden C, Takahashi A, Obi Y, Yamamoto R, Takabatake Y, Kaimori JY, Moriyama T, Horio M, Matsui I, Isaka Y.** Proteinuria-associated renal magnesium wasting leads to hypomagnesemia: a common electrolyte abnormality in chronic kidney disease. *Nephrol Dial Transplant* 34: 1154–1162, 2019. doi:10.1093/ndt/gfy119.